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<b>(21) International Application Number:</b> PCT/CA00/00191 <b>(22) International Filing Date:</b> 25 February 2000 (25.02.00) <b>(30) Priority Data:</b> 60/122,109 26 February 1999 (26.02.99) US <b>(71)(72) Applicant and Inventor:</b> LEVY, Gary [CA/CA]; 260 Henderson Avenue, Thornhill, Ontario L3T 2M2 (CA). <b>(74) Agent:</b> BERESKIN & PARR; 40th floor, 40 King Street West, Toronto, Ontario M5H 3Y2 (CA).		<b>(81) Designated States:</b> AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).  <b>Published</b> <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
<b>(54) Title:</b> MODULATORS OF FGL2 PROTHROMBINASE  <b>(57) Abstract</b>  The inventor has determined that the nucleocapsid protein from a hepatitis virus can induce the prothrombinase, fgl2. The inventor has further developed that LF-A1 also induces fgl-2. This allows for the development of therapeutic methods and compositions for modulating immune coagulation. In particular, inhibitors of the N-protein or gene or LF-A1 protein gene, or LF-A1 binding site on the fgl-2 promoter may be useful in inhibiting immune coagulation caused by a virus such as a hepatitis virus.		



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## MODULATORS OF FGL2 PROTHROMBINASE

**FIELD OF THE INVENTION**

The invention relates to modulators of the prothrombinase fgl2  
5 and the use of the modulators in controlling immune coagulation.

**BACKGROUND OF THE INVENTION**

Activation of the coagulation pathways is an important part of  
immune and inflammatory reactions and is associated with bacterial and  
viral infections (e.g. endotoxin shock, viral hepatitis), glomerulonephritis  
10 (GN), cancer, a number of gastrointestinal diseases, allograft and xeno graft  
rejection and spontaneous or stress-triggered fetal loss. Immune coagulation  
is mediated by a number of coagulants that, when triggered, activate specific  
ligands resulting in cleavage and activation of coagulation pathways that  
lead to fibrin deposition. The molecular events leading to expression of  
15 immune coagulants involve natural antibodies binding both to antigens on  
endothelial cells and Fc receptors on macrophages and endothelial cells. An  
additional mechanism is immune complex-mediated induction of  
macrophage procoagulants. These events lead to thrombin production  
which initiates platelet activation and ultimately fibrin deposition.

20 In 50% of hepatitis patients moderate to severe consumptive  
coagulopathy, or disseminated intravascular coagulopathy is found  
associated with fulminant hepatitis. Thrombi formation is observed around  
necrotic areas (Sinclair et al., 1990 and Lee, W. M., 1993). As a consequence of  
hepatitis, levels of factors II, V, VII, and X are decreased in the liver,  
25 reflecting both consumptive coagulopathy and a decrease in hepatic synthetic  
function. Also, the levels of thrombin-antithrombin complexes are high  
and platelet counts are low (Lee, W. M., 1993). These results indicate that the  
host immune system, including the coagulation pathway, is disrupted as a  
result of HBV infection. The limited host range of HBV and the difficulty to

propagate the virus in tissue culture have hampered the understanding of HBV and hepatitis B.

Murine Coronavirus infection is a model for studying acute and chronic hepatitis of humans. MHV-3 infection in BALB/cJ mice causes  
5 fulminant hepatic failure (FHF), which is characterized by macrophage activation and marked production of pro-inflammatory mediators. Especially intriguing is the ability of MHV-3 to induce de-novo synthesis of a unique procoagulant, the fgl2 prothrombinase, encoded by the fgl2 gene located on mouse chromosome 5 (Parr et al., 1995; Ning et al., 1998). Several  
10 lines of evidence implicate expression of this gene product in the pathogenesis of fulminant murine hepatitis. Firstly, levels of this prothrombinase activity correlates with the severity of the disease (Levy et al., 1983; Macphee et al., 1985). Secondly, treatment of mice with a neutralizing monoclonal antibody to the MHV-3-induced prothrombinase  
15 prevents the lethality associated with MHV-3 infection (Li et al., 1992). Concordant with these observations, expression of fgl-2 prothrombinase in liver accounts for widespread fibrin deposition in hepatic blood vessels and hepatocellular necrosis (Ding et al., 1997).

The MHV genome is a single-stranded non-segmented RNA of  
20 approximately 32 Kb (Lai and Stohlman, 1978; Pachuk et al., 1989). The RNA genome contains 7-8 genes, encoding 3-4 structural proteins and 4 nonstructural proteins (Lai and Cavanagh, 1997). An important aspect of MHV biology is the high frequency of RNA-RNA recombination between strains of MHV (Makino et al., 1989; Keck et al., 1997; Makino et al., 1986).  
25 RNA recombination may contribute to viral pathogenesis and also provides a useful tool for the study of genetic control of the biologic properties of viruses. Studies using recombinant viruses derived from MHV-JHM and MHV-A59 have demonstrated that the 3'-portion (about 25%) of the viral genome, representing RNA genomic regions encoding for all of the  
30 structural proteins, controls biologic properties such as organotropism of the



virus, the pattern of the virus-induced central nervous system pathology in mice, plaque morphology, and virus yield in tissue culture (Lavi et al., 1990; Masters and Sturman, 1990).

In view of the foregoing, there is a need to identify and  
5 characterize the molecular basis for induction of fgl2 in immune and inflammatory reactions such as in hepatitis caused BY MHV-3.

### SUMMARY OF THE INVENTION

The present inventor has shown that the nucleocapsid protein (N-protein) of mouse hepatitis virus (MHV) induces the transcription of the  
10 fgl-2 prothrombinase gene in those strains of MHV which induce fulminant hepatic failure. In particular, the inventor has shown that domain I of the MHV N-protein is required for fgl-2 induction.

Accordingly, the present invention provides a method of inducing immune coagulation comprising administering an effective  
15 amount of an nucleocapsid protein or gene or a fragment or analog thereof to an animal in need thereof. In a preferred embodiment, the N-protein comprises domain I of the MHV N-protein sequence or of the corresponding domain of the pathogenic factor of human hepatitis virus which is known to induce FGL-2, i.e., Hepatitis B and C virus. [Levy, G et al, AASLD, Nov.  
20 1999, Dallas Texas and *HEpatology*, Oct. 1999]

The present invention also includes a method of reducing or preventing immune coagulation comprising administering an effective amount of an inhibitor to the nucleocapsid protein or gene to an animal in need thereof.

25 Inhibitors of the N protein include antibodies to the N-protein. Inhibitors of the N-gene include antisense oligonucleotides to the N gene.

The present inventor has also identified that region -372 to -306 of the fgl-2 promoter is the region that is responsive to induction by the N-protein. More, particularly, in a further embodiment, the inventor has  
30 further determined that a CIS acting regulatory element, liver factor A1

binding element(LF-A1), found in the fgl2 promoter region at nucleotides -332 to -325 is responsible for fgl-2 induction. The inventor also determined that a probe specific for the LF-A1 binding element binds to the host cell LF-A1 transcription factor protein in the nucleus of a cell induced by strains of  
5 mhv known to induce fgl-2, such as, MHV-3 and MHV-A59.

Accordingly, the present invention also includes a method of reducing or preventing immune coagulation comprising administering an effective amount of an inhibitor to the promoter region of fgl2 to an animal in need thereof. Preferably, the inhibitor is an antisense oligonucleotide  
10 complimentary to nucleotides -372 to -306 of the fgl-2 gene sequence. [Genbank Accession Number M15761 and PCT/CA98/00475]

The present invention further includes a method of reducing or preventing immune coagulation comprising administering an effective amount of an inhibitor to LF-A1 to an animal in need thereof.

15 Other features and advantages of the present invention will become apparent from the following detailed description. It should be understood, however, that the detailed description and the specific examples while indicating preferred embodiments of the invention are given by way of illustration only, since various changes and modifications within the  
20 spirit and scope of the invention will become apparent to those skilled in the art from this detailed description.

#### **BRIEF DESCRIPTION OF THE DRAWINGS**

The invention will now be described in relation to the drawings in which:

25 Figure 1 is a schematic representation of the oligonucleotide maps of the recombinant viruses.

Figure 2 is a schematic and nucleic acid sequence of the fgl2 gene.

30 Figures 3A and 3B are bar graphs showing induction of fgl2 in the presence of various viruses.

Figure 4 shows the expression of fgl2 in macrophages infected with various viruses.

Figure 5 is a Western blot showing the expression of the N-protein in transfected cells.

5 Figure 6 is a bar graph showing the effect of the N-protein on the fgl2 promoter.

Figure 7A is a bar graph showing transient expression of luciferase activity by deletion constructs of the fgl2 promoter.

10 Figure 7B is a schematic of the putative regulatory elements in the fgl2 promoter responsive to N protein.

Figure 8 is a bar graph illustrating that it is the N-protein and not the I-protein of MHV259 which induces fgl-2 expression.

Figure 9 a bar graph illustrating that domain I of MHV259 is responsible for inducing fgl-2 expression.

15 Figure 10 is a confocus microscope immunofluorescence assay illustrating that N-protein localizes in the nucleus of infected macrophages.

Figure 11 is an Electrophoresis Mobility Shift DNA-protein banding assay illustrating that LF-A1 and IE1.2, but not GMSCF oligonucleotide probes bind to a transcription factor protein expressed in  
20 nuclear extracts of MHV infected cells.

Figure 12 is a bar graph illustrating that LFA-1 promoter region of fgl-2 is responsive to the N-protein of MHV.

Figure 13 is a Western blot illustrating that LFA-1 (HNF-4) is expressed in macrophages.

## 25 DETAILED DESCRIPTION OF THE INVENTION

As hereinbefore mentioned, the present inventors have determined that the nucleocapsid protein of mouse hepatitis virus (MHV) induces the transcription of the prothrombinase, fgl-2. In particular, using a set of parental and recombinant MHV strains, it was demonstrated that the  
30 nucleocapsid protein (N) of MHV induces transcription of the fgl-2

prothrombinase gene in those strains of MHV which induce fulminant hepatic failure. Two deletions found at coding sites 111-123 and 1143-1145 of structural domains I and III respectively, of the N gene may account for the important phenotypic differences observed between pathogenic and non-pathogenic strains. The present inventor has also identified that region -372 TO -306 of the fgl-2 promoter is the region that is responsive to induction by the n-protein. The inventor has further determined that a CIS acting regulatory element, liver factor A1 binding element(LF-A1), found in the fgl2 promoter region at nucleotides -332 to -325 is responsible for fgl-2 induction. The inventor also determined that the LF-A1 protein binds to the LF-AI CIS binding element of the fgl-2 promoter region. [Elizabeth M. Hardon, Monique Frain, Giacomo Paonessa and Riccardo Cortese. Two distinct factors interact with the promoter regions of several liver-specific genes. The Embo Journal Vol. 7 No. 6 pp.1711-1719, 1988]

The above finding by the inventors allows the development of therapeutic methods and compositions for modulating immune coagulation, for example, for inhibiting immune coagulation caused by viral hepatitis infection.

#### (A) METHODS OF INHIBITING IMMUNE COAGULATION

##### (1) INHIBITING THE N-PROTEIN OR N-GENE [NEW HEADER]

In one aspect, the present invention provides a method of preventing or reducing immune coagulation caused by a virus comprising administering an effective amount of an inhibitor of a nucleocapsid gene or nucleocapsid protein to an animal in need thereof.

The term "nucleocapsid" gene or protein means a nucleocapsid gene or protein from a virus and is synonymous with "capsid" or "core" gene or protein and is generally abbreviated as N-gene or N-protein. The virus can be any virus, including but not limited to, hepatitis B, C, E or G; murine hepatitis virus; human immunodeficiency virus; T-cell leukemia virus; Hanloan virus and Seoul virus.

The term "effective amount" as used herein means an amount effective and at dosages and for periods of time necessary to achieve the desired result.

The term "animal" means any member of the animal kingdom  
5 including all mammals, preferably humans.

The term "fgl-2" means any member of the fgl-2 family, including murine fgl-2 (mfgl-2) and human fgl-2(hfgl-2).

In one embodiment, the present invention provides a method of preventing or treating hepatitis comprising administering an effective  
10 amount of an inhibitor of a nucleocapsid gene or nucleocapsid protein to an animal in need thereof. preferably, the inhibitor binds to domain I of the hepatitis virus which is capable of inducing fgl-2.

**(i) Antibodies**

Polyclonal and monoclonal antibodies that bind to and  
15 neutralize the N-protein can be used to inhibit the N-protein.

Accordingly, the present invention provides a method of reducing or preventing immune coagulation caused by a virus comprising administering an effective amount of an antibody to an N-protein, or more preferably to domain I of the N-protein, to an animal in need thereof.

20 Antibodies that bind an N-protein can be prepared using techniques known in the art such as those described by Kohler and Milstein, Nature 256, 495 (1975) and in U.S. Patent Nos. RE 32,011, 4,902,614, 4,543,439, and 4,411,993 which are incorporated herein by reference. (See also Monoclonal Antibodies, Hybridomas: A New Dimension in Biological  
25 Analyses, Plenum Press, Kennett, McKearn, and Bechtol (eds.), 1980, and Antibodies: A Laboratory Manual, Harlow and Lane (eds.), Cold Spring Harbor Laboratory Press, 1988, which are also incorporated herein by reference).

30 Within the context of the present invention, antibodies are understood to include monoclonal antibodies, polyclonal antibodies,

antibody fragments (e.g., Fab, and F(ab')<sub>2</sub>) and recombinantly produced binding partners. Antibodies are understood to be reactive against the protein encoded by the nucleic acid molecule of the invention if they bind to N-protein with an affinity of greater than or equal to 10<sup>-6</sup> M. As will be appreciated by one of ordinary skill in the art, antibodies may be developed which not only bind to the protein, but which bind to a regulator of the protein, and which also block the biological activity of the protein.

Polyclonal antibodies may be readily generated by one of ordinary skill in the art from a variety of warm-blooded animals such as horses, cows, various fowl, rabbits, mice, or rats. Briefly, a N-protein of the invention or portions thereof, may be used to immunize an animal. An animal may be immunized through intraperitoneal, intramuscular, intraocular, or subcutaneous injections, in conjunction with an adjuvant such as Freund's complete or incomplete adjuvant. Following several booster immunizations, samples of serum are collected and tested for reactivity to the protein. Particularly preferred polyclonal antisera will give a signal on one of these assays that is at least three times greater than background. Once the titer of the animal has reached a plateau in terms of its reactivity to the protein, larger quantities of antisera may be readily obtained either by weekly bleedings, or by exsanguinating the animal.

Monoclonal antibodies may also be readily generated using conventional techniques as described herein. Generally, hybridoma cell lines are prepared by a process involving the fusion under appropriate conditions of an immortalizing cell line and spleen cells from an animal appropriately immunized to produce the desired antibody. Immortalizing cell lines may be murine in origin however, cell lines of other mammalian species may be employed including those of rat, bovine, canine, human origin, and the like. The immortalizing cell lines are most often of tumor origin, particularly myeloma cells but may also include normal cells

transformed with, for example, Epstein Barr Virus. Any immortalizing cell may be used to prepare the hybridomas of the present invention.

Antibody producing cells may be employed as fusion partners such as spleen cells or peripheral blood lymphocytes. The animal from  
5 which the cells are to be derived may be immunized at intervals with peptides derived from N-protein.

The immortalizing cells and lymphoid cells may be fused to form hybridomas according to standard and well-known techniques employing polyethylene glycol as a fusing agent. Alternatively, fusion may  
10 be accomplished by electrofusion.

Hybridomas are screened for appropriate monoclonal antibody secretion by assaying the supernatant or protein purified from the ascites for reactivity using the method described herein. The hybridomas are screened for antibodies which have the desired properties e.g. neutralize the N-  
15 protein and inhibit the prothrombinase activity of Fg12.

The monoclonal antibodies produced by the hybridoma cell lines of the invention are also part of the present invention. It is understood that immunoglobulins may exist in acidic, basic, or neutral form depending on their amino acid composition and environment, and they may be found  
20 in association with other molecules such as saccharides or lipids. The monoclonal antibodies produced by hybridoma cell lines of the invention may be directed against one or more of epitopes of N-protein. Any characteristic epitope associated with N-protein may provide the requisite antigenic determinant. It is contemplated that monoclonal antibodies  
25 produced by the hybridoma cell lines fall within the scope of the present invention so long as they remain capable of selectively reacting with peptides from N-protein.

The antigens recognized by the monoclonal antibodies described herein are also a part of the present invention. An antigen  
30 recognized by a monoclonal antibody produced by a hybridoma cell line of

the invention, may be localized to specific cells and tissues using conventional immunocytochemistry methods. Cryostat sections may be incubated with a monoclonal antibody of the invention and processed by the avidin-biotin-peroxidase technique (ABC Vectastain). This will  
5 determine which class of cells express an antigen of N-protein.

The present invention includes recombinant or chimeric antibody molecules. Such antibodies or binding partners may be constructed utilizing recombinant DNA techniques to incorporate the variable regions of a gene which encodes a specifically binding antibody. Within one  
10 embodiment, the genes which encode the variable region from a hybridoma producing a monoclonal antibody of interest are amplified using nucleotide primers for the variable region. These primers may be synthesized by one of ordinary skill in the art, or may be purchased from commercially available sources. Primers for mouse and human variable regions including, among  
15 others, primers for  $V_{Ha}$ ,  $V_{Hb}$ ,  $V_{Hc}$ ,  $V_{Hd}$ ,  $C_{H1}$ ,  $V_L$  and  $C_L$  regions are available from Stratacyte (La Jolla, Calif). These primers may be utilized to amplify heavy or light chain variable regions, which may then be inserted into vectors such as ImmunoZAP H or ImmunoZAP L (Stratacyte), respectively. These vectors may then be introduced into E. coli for expression. Utilizing  
20 these techniques, large amounts of a single-chain protein containing a fusion of the VH and VL domains may be produced (See Bird et al., Science 242:423-426, 1988). In addition, such techniques may be utilized to produce a "human" antibody, without altering the binding specificity of the antibody.

**(ii) Antisense molecules**

25 Antisense oligonucleotides that are complimentary to a nucleic acid sequence from a N-protein gene can also be used in the methods of the present invention to inhibit N-gene activity.

Accordingly, the present invention provides a method of preventing or reducing immune coagulation caused by a virus comprising  
30 administering an effective amount of an antisense oligonucleotide that is



complimentary to a nucleic acid sequence from an N-gene to an animal in need thereof.

The term "antisense oligonucleotide" as used herein means a nucleotide sequence that is complimentary to its target.

5 The term "oligonucleotide" refers to an oligomer or polymer of nucleotide or nucleoside monomers consisting of naturally occurring bases, sugars, and intersugar (backbone) linkages. The term also includes modified or substituted oligomers comprising non-naturally occurring monomers or portions thereof, which function similarly. Such modified or substituted  
10 oligonucleotides may be preferred over naturally occurring forms because of properties such as enhanced cellular uptake, or increased stability in the presence of nucleases. The term also includes chimeric oligonucleotides which contain two or more chemically distinct regions. For example, chimeric oligonucleotides may contain at least one region of modified  
15 nucleotides that confer beneficial properties (e.g. increased nuclease resistance, increased uptake into cells), or two or more oligonucleotides of the invention may be joined to form a chimeric oligonucleotide.

The antisense oligonucleotides of the present invention may be ribonucleic or deoxyribonucleic acids and may contain naturally occurring  
20 bases including adenine, guanine, cytosine, thymidine and uracil. The oligonucleotides may also contain modified bases such as xanthine, hypoxanthine, 2-aminoadenine, 6-methyl, 2-propyl and other alkyl adenines, 5-halo uracil, 5-halo cytosine, 6-aza uracil, 6-aza cytosine and 6-aza thymine, pseudo uracil, 4-thiouracil, 8-halo adenine, 8-aminoadenine, 8-thiol adenine,  
25 8-thiolalkyl adenines, 8-hydroxyl adenine and other 8-substituted adenines, 8-halo guanines, 8-amino guanine, 8-thiol guanine, 8-thiolalkyl guanines, 8-hydroxyl guanine and other 8-substituted guanines, other aza and deaza uracils, thymidines, cytosines, adenines, or guanines, 5-trifluoromethyl uracil and 5-trifluoro cytosine.

Other antisense oligonucleotides of the invention may contain modified phosphorous, oxygen heteroatoms in the phosphate backbone, short chain alkyl or cycloalkyl intersugar linkages or short chain heteroatomic or heterocyclic intersugar linkages. For example, the antisense oligonucleotides may contain phosphorothioates, phosphotriesters, methyl phosphonates, and phosphorodithioates. In an embodiment of the invention there are phosphorothioate bonds links between the four to six 3'-terminus bases. In another embodiment phosphorothioate bonds link all the nucleotides.

10 The antisense oligonucleotides of the invention may also comprise nucleotide analogs that may be better suited as therapeutic or experimental reagents. An example of an oligonucleotide analogue is a peptide nucleic acid (PNA) wherein the deoxyribose (or ribose) phosphate backbone in the DNA (or RNA), is replaced with a polyamide backbone  
15 which is similar to that found in peptides (P.E. Nielsen, et al Science 1991, 254, 1497). PNA analogues have been shown to be resistant to degradation by enzymes and to have extended lives *in vivo* and *in vitro*. PNAs also bind stronger to a complimentary DNA sequence due to the lack of charge repulsion between the PNA strand and the DNA strand. Other  
20 oligonucleotides may contain nucleotides containing polymer backbones, cyclic backbones, or acyclic backbones. For example, the nucleotides may have morpholino backbone structures (U.S. Pat. No1 5,034, 506). Oligonucleotides may also contain groups such as reporter groups, a group for improving the pharmacokinetic properties of an oligonucleotide, or a  
25 group for improving the pharmacodynamic properties of an antisense oligonucleotide. Antisense oligonucleotides may also have sugar mimetics.

The antisense nucleic acid molecules may be constructed using chemical synthesis and enzymatic ligation reactions using procedures known in the art. The antisense nucleic acid molecules of the invention or a  
30 fragment thereof, may be chemically synthesized using naturally occurring

nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed with mRNA or the native gene e.g. phosphorothioate derivatives and acridine substituted nucleotides. The antisense sequences  
5 may be produced biologically using an expression vector introduced into cells in the form of a recombinant plasmid, phagemid or attenuated virus in which antisense sequences are produced under the control of a high efficiency regulatory region, the activity of which may be determined by the cell type into which the vector is introduced.

10 The antisense oligonucleotides may be introduced into tissues or cells using techniques in the art including vectors (retroviral vectors, adenoviral vectors and DNA virus vectors) or physical techniques such as microinjection. The antisense oligonucleotides may be directly administered *in vivo* or may be used to transfect cells *in vitro* which are then  
15 administered *in vivo*. In one embodiment, the antisense oligonucleotide may be delivered to macrophages and/or endothelial cells in a liposome formulation.

### (iii) Other N-protein inhibitors

In addition to antibodies and antisense oligonucleotides, other  
20 substances that inhibit N-protein may be isolated. N-protein binding peptides may be isolated by assaying a sample for peptides that bind to the N-protein or domain I of the N-protein. Any assay system or testing method that detects protein-protein interactions may be used including co-immunoprecipitation, crosslinking and co-purification through gradients or  
25 chromatographic columns may be used. Biological samples and commercially available libraries may be tested for N-protein binding peptides. For example, labelled N-protein may be used to probe phage display libraries. In addition, antibodies prepared to the peptides of the invention may be used to isolate other peptides with N-protein binding

affinity. For example, labelled antibodies may be used to probe phage display libraries or biological samples.

Additionally, a DNA sequence encoding a N-protein may be used to probe biological samples or libraries for nucleic acids that encode N-protein-binding proteins.

## (2) INHIBITING LF-A1

In another aspect, the present invention provides a method of reducing or preventing immune coagulation comprising administering an effective amount of an inhibitor to LF-A1 to an animal in need thereof.

Inhibitors of LF-A1 include antibodies and antisense molecules to LF-A1 that may be prepared using standard techniques as described above for the N-protein and gene. In addition other molecules that inhibit LF-A1 may be isolated using the methods described above for isolating inhibitors of the N-protein or gene.

## (3) INHIBITING FGL-2 PROMOTER REGION

In a further aspect, the present invention provides a method of reducing or preventing immune coagulation comprising administering an effective amount of an inhibitor to the promoter region of fgl-2 to an animal in need thereof. Preferably, the inhibitor is an antisense oligonucleotide complimentary to nucleotides -372 TO -306 , or preferably -332 to -325, of the fgl-2 Gene sequence. (Genbank Accession Number m15761 and PCT/CA98/00475).

## (B) METHODS OF INDUCING IMMUNE COAGULATION

In an alternate embodiment, the present invention includes methods of inducing immune coagulation by administering an N-protein or gene. Methods that induce immune coagulation may be useful in treating conditions which require an increase in coagulant activity. Such methods can also be used to induce fetal loss.

Accordingly, the present invention provides a method of inducing immune coagulation comprising administering an effective

amount of an N-protein or gene or a fragment or analog thereof to an animal in need thereof.

As used herein, a fragment of a N-protein or gene means any portion of the full length gene or protein that is sufficient to induce fgl-2 transcription or activity. Preferably, the fragment comprises domain I of the N-gene which includes nucleotides 400-500 of the N-gene sequence. [Genbank Accession number M35156]

As used herein, an analog (or mimetic) of the N-gene or protein means any molecule that is functionally similar to the N-protein or gene in that it can also induce fgl-2 transcription or activity. Analogs or mimetics of the n-protein or gene that may be useful in the present invention may be identified using methods known in the art.

In a preferred embodiment, the n-protein analog is a structural as well as functional analog which includes any peptide having an amino acid residue sequence substantially identical to the N-protein in which one or more residues have been conservatively substituted with a functionally similar residue and which displays the ability to mimic the N-protein in inducing fgl-2 activity. Examples of conservative substitutions include the substitution of one non-polar (hydrophobic) residue such as alanine, isoleucine, valine, leucine or methionine for another, the substitution of one polar (hydrophilic) residue for another such as between arginine and lysine, between glutamine and asparagine, between glycine and serine, the substitution of one basic residue such as lysine, arginine or histidine for another, or the substitution of one acidic residue, such as aspartic acid or glutamic acid for another. The phrase "conservative substitution" also includes the use of a chemically derivatized residue in place of a non-derivatized residue provided that such polypeptide displays the requisite activity.

Structural analogs of the N-protein may be identified by probing biological samples and commercially available peptide libraries with an

antibody that binds to the N-protein as described herein above. For example, labelled antibodies to N-protein may be used to probe phage display libraries. Any assay system or testing method that detects protein-protein interactions may be used to measure the binding of the antibody to a peptide  
5 in the sample including co-immunoprecipitation, crosslinking and co-purification through gradients or chromatographic columns may be used.

The structure of the N-Protein may also be used to identify lead compounds for drug development for use in methods of inducing immune coagulation. A comparison of the structures of peptides similar in sequence  
10 to the N-protein, but differing in the biological activities they elicit in target fgl-2 molecules can provide information about the structure-activity relationship of the induction of fgl-2. Information obtained from the examination of structure-activity relationships can be used to design either modified peptides, or other small molecules or lead compounds which can  
15 be tested for the induction of fgl-2. The structure of the peptides identified can be readily determined by a number of methods such as NMR and X-ray crystallography.

Fragments and analogs and structural analogs for the LF-A1 protein shall have a corresponding meaning.

## 20 (C) COMPOSITIONS

The antibodies, antisense oligonucleotides or other inhibitors of N-protein identified using the methods described herein as well as the N-protein and nucleic acid sequences including fragments and analogs, may be incorporated into a pharmaceutical composition containing the substance,  
25 alone or together with other active substances.

In one aspect, the present invention provides a composition for use in inhibiting procoagulant activity in an animal comprising (a) an antibody specific for a N-protein; (b) antisense nucleic acid molecules

complimentary to N-protein; or (c) an inhibitor identified using the methods as described above in admixture with a suitable diluent or carrier.

5 In another aspect, the present invention provides a composition for use in inducing procoagulant activity in an animal comprising a nucleic acid sequence encoding N-protein or an N-protein or a fragment or analog of the n-protein or the nucleic acid encoding the N-protein in admixture with a suitable diluent or carrier.

10 In another embodiment the present invention provides a composition for use in inducing procoagulant activity comprising an effective amount of LF-A1 protein or fragment or analog thereof in admixture with a suitable diluent or carrier.

15 In another aspect the present invention provides a composition for suppressing or inhibiting procoagulant activity comprising an effective amount of an inhibitor of LF-A1 in admixture with a suitable diluent or carrier. Such an inhibitor can include but is not limited to an antibody to LF-A1 or to a molecule which blocks the binding of LF-A1 to the LF-A1 CIS binding element of the fgl-2 promoter, such as an anti-sense oligonucleotide to the LF-A1 binding element of the promoter region of fgl-2.

20 Such pharmaceutical compositions can be for oral, topical, rectal, parenteral, local, inhalant or subcutaneous, intradermal, intramuscular, intathecal, vaginal, transperitoneal, placental and intracerebral use. They can be in liquid, solid or semisolid form, for example pills, tablets, creams, gelatin capsules, capsules, suppositories, soft gelatin capsules, gels, membranes, tubelets, solutions or suspensions.

25 The pharmaceutical compositions of the invention can be intended for administration to humans or animals. Dosages to be administered depend on individual needs, on the desired effect and on the chosen route of administration.

The pharmaceutical compositions can be prepared by per se known methods for the preparation of pharmaceutically acceptable compositions which can be administered to patients, and such that an effective quantity of the active substance is combined in a mixture with a pharmaceutically acceptable vehicle. Suitable vehicles are described, for example, in Remington's Pharmaceutical Sciences (Remington's Pharmaceutical Sciences, Mack Publishing Company, Easton, Pa., USA 1985).

On this basis, the pharmaceutical compositions include, albeit not exclusively, the active compound or substance in association with one or more pharmaceutically acceptable vehicles or diluents, and contained in buffered solutions with a suitable pH and iso-osmotic with the physiological fluids. The pharmaceutical compositions may additionally contain other agents such as adjuvants to enhance immune responsiveness.

The antisense nucleic acid molecules of the invention may be used in gene therapy to inhibit immune procoagulant activity in animal with a viral infection. Recombinant molecules comprising an antisense sequence or oligonucleotide fragment thereof, may be directly introduced into cells or tissues *in vivo* using delivery vehicles such as retroviral vectors, adenoviral vectors and DNA virus vectors. They may also be introduced into cells *in vivo* using physical techniques such as microinjection and electroporation or chemical methods such as coprecipitation and incorporation of DNA into liposomes. Recombinant molecules may also be delivered in the form of an aerosol or by lavage. The antisense nucleic acid molecules of the invention may also be applied extracellularly such as by direct injection into cells.

#### (D) Vaccines

The present invention also contemplates a vaccine against a virus causing immune coagulation comprising an amount of an N-protein or peptide which is effective to induce an immune response against N-protein.



In one embodiment, the present invention provides a vaccine for treating or preventing viral hepatitis comprising an effective amount of an N-protein or peptide in admixture with a suitable diluent or carrier.

The vaccine may be a multivalent vaccine and additionally  
5 contain immunogens related to other diseases in a prophylactically or therapeutically effective manner.

The vaccine may also comprise an immunologically acceptable carrier such as aqueous diluents, suspending aids, buffers, excipients, and one or more adjuvants known in the art. Examples of adjuvants include the  
10 lipid A portion of gram negative bacteria endotoxin, trehalose dimycolate of mycobacteria, the phospholipid lysoleathin, dimethyl-ditadecyl ammonium bromide (DDA), linear polyoxypropylene-polyoxyethylene (POP-POE) block polymers and liposomes. The vaccine may also contain cytokines that can enhance the immune response including GM-CSF, IL-2, IL-12, TNF and  
15 IFN $\gamma$ . The vaccine may also contain preservatives such as sodium azide, thimersol, beta propiolactone, and binary ethyleneimine.

The vaccines of the invention can be intended for administration to animals, including mammals, avian species, and fish; preferably humans and various other mammals, including bovines, equines,  
20 and swine.

The vaccines of the invention may be administered in a convenient manner, such as intravenously, intramuscularly, subcutaneously, intraperitoneally, intranasally or orally. The dosage will depend on the nature of the disease, on the desired effect and on the chosen  
25 route of administration, and other factors known to persons skilled in the art.

A vaccine prepared using the methods described herein may be tested in *in vivo* animal systems to confirm their efficacy in the prophylaxis or active immunization and treatment of the relevant disease and to  
30 determine appropriate dosages and routes of administration.

- 20 -

The present invention also includes the use of the antibodies that bind the N-proteins and portions thereof of the invention as a means of passive immunization.

The present invention also includes DNA immunization with  
5 an N-protein gene or portion thereof.

The following non-limiting examples are illustrative of the present invention:

### EXAMPLES

#### Example 1

#### 10 INDUCTION OF FGL2 BY N-GENE OF MHV-3

##### Materials and Methods

##### Mice

Female Balb/cJ mice, 6-8 weeks of age, from Charles River Laboratories (St. Constant, Quebec) were kept in microisolated cages and  
15 housed in the animal facilities at the Toronto Hospital, and fed a standard Lab chow diet and water ad libitum.

##### Virus

MHV-3 was obtained from American Type Culture Collection [(ATCC), Rockville, MD] and plaque purified on monolayers of DBT cells.  
20 Parental viruses A59, JHM, MHV-2 and 2 sets of recombinant viruses have been described previously (Lai, 1992). The schematic representations of oligonucleotide maps of the recombinant viruses are presented in Figure 1 (Lai, 1992). To ensure that the recombinant virus strains are clonal, ML3 and ML11 were plaque purified 3 times in 17Cl-1 cells. The purified strains were  
25 used for creating N gene expression constructs and transfection experiments.

##### Cells

Peritoneal macrophages were harvested from Balb/cJ mice 4 days after intraperitoneal administration of 1.5 ml of 3% thioglycolate (Difco Laboratories, Detroit, Michigan) as previously described (Ding et al., 1997).  
30 Macrophages were resuspended in RPMI 1640 (ICN Biomedicals Inc., Costa

Mesa, California) supplemented with 2 mM glutamine (Sigma Chemical Co., St. Louis, MO) and 2% heat-inactivated fetal calf serum (Flow Laboratories, Mississauga, Ontario, Canada). Macrophages were greater than 95% in purity as determined by morphology and non-specific esterase stain. Viability exceeded 95% by trypan blue exclusion. Chinese hamster ovary cell line (CHO cells) were from ATCC.

#### PCA

MHV and recombinant virus-infected macrophages, at a multiplicity of infection (MOI) of 2.5, were incubated for 8h in RPMI 1640 supplemented with 10% Fetal Bovine Serum and 200 mM glutamine. Mock-infected macrophages and MHV-3-infected macrophages represented negative and positive controls, respectively. Macrophages were evaluated for functional PCA in a one-stage clotting assay, as previously described (Levy et al., 1981). Following incubation, samples to be assayed for PCA were washed three times with unsupplemented RPMI 1640 and resuspended at a concentration of  $10^6$ /ml. Samples were assayed for the ability to shorten the spontaneous clotting time of normal citrated human platelet-poor plasma. Milliunits of PCA were assigned by reference to a standard curve generated with serial log dilutions of a standard rabbit brain thromboplastin (Dade Division, American Hospital Supply Co., Miami, Florida).

#### RT-PCR

Expression of fgl2 was detected by RT-PCR. Freshly isolated macrophages, at a multiplicity of infection (MOI) of 2.5, were infected with different strains of viruses for 6 hours.  $1 \times 10^7$  macrophages were pelleted in 1.5 ml Eppendorf tube and total cellular RNA was isolated by 8M acid-guanidium hydrochloride extraction in a modified procedure as previously described (Evans and Kamdar, 1990). The quantity and quality of RNA was examined by spectrophotometry and on a 1% analytical agarose gel containing formaldehyde. RNA (5 $\mu$ g) was reverse transcribed using Moloney Murine Leukemia Virus Reverse Transcriptase (M-MLVRT) in 20

µl reactions, as recommended by manufacturer. PCR was then performed in 50 µl reactions using 1µl portion of cDNA and the primers fgl2-318 (TGC CCA CGC TGA CCA TCC A) corresponding to nucleotide 318 ñ 336 of Balb/c fgl2 cDNA (M 15761 ) and FGL2-1224 ( GAG ACA ACG ATC GGT ACC CCT) corresponding to nucleotides 1224 -1244 of Balb/c fgl2 cDNA. (M16238), which yield a 906 bp band in 1% agarose DNA gel. Amplification products were not obtained when reverse transcriptase was omitted (data not shown). RT-PCR for GAPDH was also set up as an internal control to assess the quality of first strand synthesis.

#### 10 Creation of N gene and fgl2 promoter constructs

Restriction enzymes used to create constructs were obtained from GIBCO BRL, Life Technologies, USA. All plasmids were purified using Qiagen Maxiprep Kits, and grown in DH5 E. coli bacteria (GIBCO BRL).

MHV-2 and A59 are different sizes. The entire N gene coding regions and 3' UTRs of MHV-A59, MHV-2, ML3 and ML11 were amplified by RT-PCR. RNA was originally extracted from infected macrophages. The sense primer ACG ATG TCT TTT GTT CCT GGG was phosphorylated chemically at position 1 to achieve directed insertion and ligation of PCR products to its vector; the anti sense primer at position 1654 TTT TTT TTT GTG ATT CTT CCA had a poly T group to match the poly A tail at the 3' end of nucleocapsid genomic RNA. The N gene fragments were subcloned into the 5.0 Kb expression vector pCR 3.1 (Invitrogen), under the control of the CMV promoter and bovine growth hormone 3'-processing signals. External restriction endonuclease Hind III and Pst I and internal restriction endonuclease Eco RI, Eco RV were used to analysis the size and orientation of N gene insert in recombinant plasmid constructs.

Luciferase Reporter Constructs: A 1.3 kb DNA fragment flanking the 5' end of mouse fgl2 was released by restriction digestion with EcoRV and Sal I from a subclone pBluescript-m166 (pm166) of mouse genomic P1 plasmid (Genome System Inc.) which contains the entire mouse

fgl2 gene. This fragment was sequenced by cycle sequencing on an automated DNA sequencer (Model 377, Applied Biosystems) using dideoxy dye terminator chemistry. This sequence has been deposited into Genbank with the Accession number AF025817 (Figure 2) (Koyama et al., 1987). This 1.3 kb fragment was inserted into Sma I and Xho I sites of the pGL2-basic luciferase reporter vector (Promega) to form pfgl2(-1328)LUC. 5' deletion constructs of fgl2 promoter were made by first amplifying the specific fragment using pm166 as template and then cloned into PCR 2.1 cloning vector (Stratagene) and resubcloned into pGL2-basic plasmid at Xho I and Hind III sites. The reverse primer (GCC ACA ACC AAC CAG GAA G) was used to make all deletion constructs by PCR amplification. The upstream primers used were: GAG CTG AGT GAT GGG GAA GGA for pfgl2(-693)LUC, CCA CTG ACG ATT ACA TAG CC for pfgl2(-625)LUC, GGA CCT TTG TTC TGA TTA GGG GC for pfgl2(-511)LUC, CGC AGA CAT TTA GAC GTT CC for pfgl2(-372)LUC, and GGG CAC TGG TAT TAC AAC TGT for pfgl2(-306)LUC. All promoter-luciferase report constructs were sequenced to confirm the orientation and to verify the sequence. Positive control, pGL2 Control plasmid with SV-40 promoter, and RSV  $\beta$ -gal vector were from Promega. A 2Kb tissue factor promoter construct pTF(-2Kb)LUC was a kind gift of Dr. Nigel Mackman (Mackman et al., 1990).

#### N gene Sequence of different virus strains

The N gene sequence of multiple clones for each strain of virus was determined using primer-directed strategies by cycle sequencing on an automated DNA sequencer, using the ABI PRISMTM dRhodamine Terminator Cycle Sequencing Reading Reaction Kit (Model 377, PE Applied Biosystems). The T7 primer and pCR3.1 reverse primer were used for 5' to 3' and 3' to 5' sequence, respectively. A new primer CTC AGG GCT TTT ATG TTG AAG (MHV-ND557) at position 557 was also designed based on the outcome of sequencing and the published cDNA sequence of MHV-A59 to complete the sequencing. Extension products were purified by

ethanol/sodium acetate precipitation. Samples were subjected to electrophoresis on the ABI PRISM 310. The sequence was analyzed using the DNAsis for windows, sequence analysis software (Hitachi Software Engineering America Ltd, San Bruno, CA, USA).

## 5 Transfection

CHO were cultured in 6-well plates until 50-80% confluence. 1  $\mu$ g of N gene construct DNA, 0.5  $\mu$ g of fgl2 promoter construct DNA and 0.25  $\mu$ g of beta-gal DNA (as a marker for transfection efficiency by beta-gal assay) in 100  $\mu$ l of OPTI-DMEM medium were mixed by vortexing with 3.5  $\mu$ l of lipofectAMINETM (2  $\mu$ g/ $\mu$ l) in 100  $\mu$ l of OPTI-DMEM medium. After incubation of the mixture at room temperature for 30 min, 1.8 ml of OPTI-DMEM medium were added to bring up the volume to 2 ml. One ml of this mixture were distributed into one of the duplicated wells with CHO cells and transfection were proposed at 37 o C with 5% CO<sub>2</sub> for 44- 48 hrs. Cells were harvested in lysis buffer and freeze-thawed 3 times in liquid N<sub>2</sub>. Aliquots of supernatants were assayed for beta-galactosidase and luciferase activity.

## Western blot

At 48 hrs post transfection with N gene and fgl2 (-1.3 Kb/+9) promoter constructs, cultures of 1 x 10<sup>6</sup> of CHO cells were collected and lysed in 100  $\mu$ l of Western blot lysis buffer with protease inhibitor. Twenty (20)  $\mu$ l of lysate were resolved by SDS-PAGE and then transferred to a nitrocellulose membrane. After blocking in 4% milk-PBS for 1 hr, membrane was probed with a monoclonal antibody against the N protein at 4°C overnight, followed by washing in 2% milk-PBS Tween a total of 5 times. The membrane was then incubated with goat anti mouse IgG labeled with horseradish peroxidase for 1 hr and washed in 2% milk-PBS Tween 5 times. Substrates luminol and enhancer were added and incubated for 1 min. The

membrane was then exposed to Kodak XAR-5 film with intensifying screens for 10 minutes.

### Statistical Analysis

Data are expressed as mean + standard deviation (SD) where applicable. Student's t test for unpaired samples (two tailed) was used to analyze the data.

#### (a) *Expression of Balb/c macrophage functional PCA induced by parental viruses and their recombinants.*

Studies were undertaken using parental A59, JHM and MHV-2 strains and 2 sets of recombinant viruses between them (A59 x JHM, A59 x MHV-2) (Figure 1). MHV-A59 infection of macrophages resulted in a marked elevation of functional procoagulant activity (PCA), similar to what was reported previously for MHV-3 (Parr et al., 1995 and Ding et al., 1997) whereas JHM and MHV-2 failed to induce PCA (Figure 3A). A59 X JHM-derived recombinants B1, RL1 and IL27, in which the 3'-portion of the genome is derived from MHV-A59, induced high level of functional PCA. In contrast, CA13 and CA43, two recombinants in which the 3'-portion of the genome is derived from MHV-JHM, did not induce PCA, suggesting that the 3'-portion of the MHV-A59 genome may contain a viral genetic determinant needed for induction of functional PCA, which is lacking in the corresponding region of MHV-JHM. To better delineate the candidate genes required for induction of fgl2, we next studied a set of recombinants derived from MHV-A59 X MHV-2, which have multiple crossovers in a single genome, particularly within the 3' portion of the genome. Recombinants ML3 and ML10 induced functional PCA, while ML11 did not (Figure 3B). The differences between recombinants ML3 and ML11 in the 3'-portion the genome maps within the 5'-end of the N gene. Therefore, the ability of recombinants to induce functional PCA parallels the presence or absence of MHV-A59 sequences representing the N gene, strongly suggesting that the MHV-A59 N gene is responsible for activation of fgl2.

(b) *Induction of fgl2 transcription by parental and recombinant viruses.*

RT-PCR analysis demonstrated that the presence or absence of detectable fgl2 mRNA in macrophages infected with various parental or recombinant viruses correlated with the results of assays for functional fgl2 prothrombinase (Figure 4).

(c) *Sequence comparison of N-genes*

The entire N gene sequence was determined for MHV-A59, MHV-2, and the recombinant viruses ML3 and ML11, as described in Materials and Methods (sequence data not shown).

A comparison of different N gene sequences is shown in Table 1. Compared to the published sequence of A59 in GeneBank (M35156), the sequence of MHV-A59 used in this study had two point mutations at nt 441 (T→A) and nt 1613 (T→C); ML3 had three point mutations at nt 405 (C→A), nt 441 (T→A), nt 1613 (T→C). Sequence differences between the N genes of MHV-A59 and MHV-2 (AF061835) were mainly within two regions corresponding to nt 400-500 (**domain I**) and nt 1100-1200 (**domain III**) of the MHV-A59 sequence. In addition, MHV-2 had a 12 nt-deletion at nt 111-123 and a 3-nt insertion at nt 1143-1145 when compared with MHV-A59. In contrast, the 3'-untranslated regions (UTRs) following the N gene are completely identical between MHV-A59 and MHV-2.

(d) *Effect of N protein on fgl2 promoter activity*

To establish that the N gene of MHV is responsible for the induction of fgl2 gene, co-transfection of CHO cells with the N gene under CMV promoter and fgl2 promoter-luciferase reporter constructs were performed. The expression of the N protein was confirmed by Western blotting (Figure 5). CHO cells co-transfected with the N gene construct from A59 and a murine fgl-2 promoter/reporter luciferase construct showed a 6-fold increase in luciferase activity compared to cells transfected only with the fgl-2 promoter/reporter luciferase construct (Figure 6). In contrast, MHV-2 N protein expression did not enhance fgl-2 expression. To confirm



the specificity of the effect of N protein on fgl2 promoter activity, pTF(-2Kb)LUC and a pGL2-control vector under SV 40 promoter were cotransfected respectively with the MHV-A59 N gene construct. There was no significant increase in luciferase expression when these 2 constructs were co-transfected respectively with or without the MHV-A59 N gene construct (Figure 6).

This was further evidenced by studying the effect of N- and I-protein [Fischer et al., Journal of Virology, Feb, 1997, p. 996-1003, vol 71, No. 2] mutations of domain I of MHV on expression of Balb/cJ macrophage functional PCA. Macrophages from Balb/cJ mice were infected with MHV-3, MHV-A59, N-mutants: MHV-2 and MHV-JHM, and I-mutants: MHV-ALB110 and MHV-ALB111 at a M.O.I. of 2D.5 for 8-10 hrs and harvested for measurement of pca activity. The results are shown in Figure 8, where values represent the mean +/- S.D. of three separate experiments done in triplicate. \* represents  $P < 0.01$  compared with unstimulated macrophages. The results indicate that MHV-3, MHV-A59 and the I-protein mutations induced Balb/cJ macrophage functional PCA, while MHV-2 does not.

To establish which portion of the n-protein was responsible for fgl-2 induction, N gene constructs from MHV-A59 and MHV-2 and a series of N gene mutants from MHV-A59 domain I (A59G12S, A59P38L, A59P38DEL, A59NQN 40-42DEL) and domain III (A50V321A AND A59E85Q) were cotransfected with the wild type fgl2 promoter-LUC respectively into cho cells. The results are shown in Figure 9, where relative luciferase activity is expressed in fold increase compared with cho cells cotransfected with n construct from MHV-2. PGL@-basic vector was used as a negative control. Values represent the mean +/- S.D. of five separate experiments done in triplicate. \* represents  $P < 0.01$  compared with cells cotransfected with MHV-2 N construct. # Represents  $P < 0.01$  compared with cells cotransfected with n gene construct from wild type MHV-A59. As can be seen by Figure 9, mutations in structural domain I did not induce fgl2-promoter-luciferase

activity, which suggests that it is domain I which is responsible for fgl-2 induction.

*(e) Mapping of the fgl2 promoter.*

In order to characterize the region in the fgl2 promoter, which  
5 is responsive to N protein of MHV-A59, constructs containing progressive deletions of the -1328 bp fragment were cotransfected with either N gene constructs or empty pCR 3.1 vector in CHO cells (Figure. 7A). Preliminary mapping of the fgl2 promoter has defined a region from -372 to -306 to be responsive to induction of N protein. The activity of the deletion construct  
10 -306 was comparable to the promoterless plasmid pGL2-basic. Using DNAsis software, three positive cis acting regulatory elements were identified within this region which included a liver factor A1 binding element (LF-A1, -332 to -325), human cytomegalovirus immediate early gene 1.2 (IE1.2, -345 to -336) regulatory elements and granulocyte macrophage colony-stimulating  
15 factor binding element (GMCSF, -353 to -346) (Figure. 7B).

In order to determine which of the cis elements of the fgl2 promoter region were involved in N-protein induced fgl2 transcription, a series of experiments were conducted.

It was shown by confocus microscope immunofluoresence that  
20 n protein is present in the nucleus of infected macrophages. In figure 10, macrophages from Balb/cJ mice were planted as a monolayer on the glass slide flask(VWR) And infected with MHV-A59 and MHV-2. Cells were stained for nucleocapsid protein in the nucleus of infected macrophages.

The next step was to determine which of the 3 CIS elements  
25 would bind to proteins expressed in the nuclear extracts of infected cells.

Electrophoresis mobility shift assays(EMSA) and competition studies were conducted, as shown in Figure 11. CHO cells were incubated with MHV-259 and then DNA-protein banding was performed using band shift kit according to the instruction (Pharmacia Biotech).

Band shift assays were performed with oligonucleotides corresponding to the LF-AL binding element (Lanes 1-4) or IE1.2 binding element (Lanes 5-8) or GMCSF binding element (Lanes 9-12) within the promoter region of fgl2 gene. The sequences of the oligonucleotides used were:

5 For the LF-A1 site (-338/-319 of fgl-2 promoter region):  
5'-CAC TAG TGG ACC AAG TAT-3' AND 5'- AAT TAT ACT  
TGG TCC ACT AGT G-3';

For the IE1.2 SITE (-353/-336 of fgl2 promoter region)):  
5'-TTC CAA CTC TTT CCC AC-3' AND 5'-AAT TGT GGG A A A  
10 GAG TTG CAA;

For the GMCSF site (-368/-351 of fgl2 promoter region):  
5'-ACA GAC ATT TAG AGG TTC-3' AND 5'-AAT TGA ACG  
TCT AAT GTC TGT-3'.

15 The probes used in EMSA were chemically synthesized  
oligonucleotides. 200NG of sense and anti sense probe were mixed in  
annealing buffer in a total of 20 uL of reaction and incubated at 65 OC for 5  
min, Then gradually cooled down to room temperature to be annealed to  
form a double strand dna probe. 2.5 uL of the double- stranded probe was  
20 labeled with 20 UCI of [(-<sup>32</sup>P)]DATP using klenow fragment (Pharmacia  
Biotech). DNA was separated from non-incorporated radioisotope by probe  
QUANT™ G-50 micro-columns.

For competition experiments, nuclear extracts (nu) were  
preincubated with either specific competitor (cold probes) or non specific  
25 competitor. After addition of radiolabel led probes, the free DNA and the  
DNA protein complex were separated on a 6% POLYACRYLAMIDE GEL.  
Lane 1,5,9: free 32P-probe; Lane 2,6,10: 32P-probe + NU; Lane 3,8,11: 32P-probe

- 30 -

+ cold specific competitor + NU; Lane 4,7,12: <sup>32</sup>P-probe + non specific competitor + NU.

Nuclear extracts were incubated with <sup>32</sup>P-labeled double-stranded oligonucleotides probe for one of the CIS ELEMENTS FOR  
5 15 min in a total of 20 uL reaction at 4 °C In the presence of 10 MM TRIS-HCL pH 7.5, 50 MM NACL, 0.5 MM DTT, 10% GLYCEROL, 0.05% NP40, 1 UG of poly (DI-DC), 50 MM NACL and 5 MM MGCL2. Protein-DNA complexes were analyzed by electrophoresis in 5% nondenaturing gels, followed by auto radiography.

10 The assay results show that LF-A1 and IE1.2 oligonucleotide probes bind to protein expressed in the nuclear extracts. The banding suggests that LF-A1 binds to the LF-A1 host transcription factor of the infected cell. The LF-A1 transcription factor has previously been described in, Elizabeth M. Hardon, Monique Frain, Giacomo Paonessa and Riccardo  
15 Cortese. "Two Distinct Factors Interact With The Promoter Regions Of Several Liver-specific Genes." The EMBO Journal Vol. 7 No.6 pp.1711-1719, 1988.

To study whether LF-A1 or IE1.2 was responsible for fgl-2 induction, the effect of mutations in these elements on transient expresion  
20 of luciferase activity of MFGL-2 was studied in response to MHV-A59 N protein in CHO cells. 0.5 Micrograms of N gene constructs from MHV-A59, MHV-2 was cotransfected with -.25 micrograms of wild-type (WT) PFGL2 (-1328)LUC or its mutant for candidate CIS elements LF-A1 (LF1MUT), IE1.2 (IE1.2MUT) or LFA1 and IE1.2 double mutation (LFAL/IE1.2MUT)  
25 respectively in CHO cells for 40-44 hrs. Cells were harvested and freeze-thawed three times for measurement of luciferase activity. PGL2-basic vector was used as a negative control. The results are shown in Figure 12, where values represent the mean ± S.D. of four separate experiments done in triplicate. \* represents P < 0.01 compared with cells cotransfected with MHV-  
30 2 N construct. # represents P <0.01 compared with cells cotransfected with

wide type PFG12 (-1328) LUC. The results show that LF-A1 is responsible for fgl2 induction.

As LF-A1 has not previously been reported to be expressed in cells other than the liver and kidney, a western blot analysis for detection of LF-A1 protein in macrophages in Balb/cJ was conducted. The results are shown in Figure 13. Nuclear proteins (NU) were extracted from macrophages infected with MHV at a multiplicity of infection (M.O.I) OF 2.5. CHO cells transfected with HNF-4 (LF-A1) expression construct were used as positive control. Twenty (20) ML of lysate were resolved by SDS-page and then transferred to a nitrocellulose membrane. Membrane was probed with a polyclonal antibody against the HNF-4 protein at 4°C overnight, followed by incubation with goat anti rabbit IGG labeled with horseradish peroxidase as described under the methods and materials section above for "western blot. Lane 1. HNF-4 expression construct + CHO NU; Lane 2. MΦ NU; Lane 3. MΦ + MHV-A59 NU; Lane 4. MΦ + MHV-2 NU. Figure 13 shows that LF-A1 IS present in macrophages.

### Summary

Studies using a model of viral hepatitis induced by infection with MHV-3 have provided significant insight into the mechanisms underlying the pathogenesis of this disease and have suggested novel approaches to therapy (Ning et al., 1998; Cattral and Levy, 1995; Sidwell et al., 1977). Furthermore, recent studies have demonstrated the role of the selective expression of the fgl2 prothrombinase in the pathogenesis of MHV-3-induced fulminant liver failure (Parr et al., 1995; Ning et al., 1998; Li et al., 1992; Ding et al., 1997).

The studies presented here demonstrate that infection of macrophages with MHV-A59 resulted in elevated PCA, a result similar to that observed during infection of macrophages with MHV-3. This contrasts with the results obtained with MHV-JHM and MHV-2, which do not induce

elevations of PCA in infected macrophages. Recombinant viruses (n=5) which derive a portion of their genomic RNA from the 3'-region of MHV-A59 induces functional PCA, whereas those containing a 3'-region derived from JHM or MHV-2 (n=5) do not, suggesting that the 3'-portion of the genome may play an important role in the induction of fgl-2 transcription. The sequence differences in the genome in the 3'-area of interest lies mainly within the 5' end of the N gene. The sequence differences in the N gene between inducers and non-inducers are a 12-nucleotide deletion at nt 111-123 of the coding region for the structural domain I and a 3- nucleotide insertion at nt 1143-1145 of the coding region for structural domain III, which may account for the inability of JHM or MHV-2 to induce fgl-2. CHO cells cotransfected with the N gene construct from A59 and with the fgl-2 promoter construct showed a 6-fold increase in luciferase activity in contrast to baseline or MHV-2-cotransfected cells.

These findings strongly suggest that the N gene is responsible for fgl-2-induction. It should be noted that induction of fgl-2 could not be explained by differences in virus replication, as MHV-2, a non-inducer, replicates to higher titers than A59 while A59 replicates to higher titers than JHM. It is also of great interest that all of the recombinants of MHV-2 X A59 have the MHV-2-derived leader sequence whereas the majority of recombinants between A59 X JHM contain the A59 leader (Keck et al., 1988). It is not clear whether the leader sequence differences among viruses are responsible for the altered growth property of the virus.

MHV N protein has been proposed to consists of three conserved structural domains (I basic, II basic, and III acidic) which are tethered to each other by two regions of variable amino acid composition (designated A and B) (Parler, M.M. and Masters, P.S., 1990). The sequence differences in the N gene between viruses which induce fgl-2, such as MHV-3 and MHV-A59, and MHV-2 lies within two regions corresponding to domain I (nt 400-500) and domain III (nt 1100-1200), respectively. The

inventor has shown that it is primarily the differences in domain I which account for inability of MHV-2 to induce the fgl2 gene.

Co-transfection of an N gene expression construct with the fgl2 promoter/reporter luciferase confirms that the N gene of A59 accounts for  
5 induction of fgl2. Possible mechanisms by which n protein induces fgl2 prothrombinase expression include the transport of the N protein into the nucleus of infected cells, acting as a transcription activator for fgl2; or modulation of signal transduction pathways that regulate host transcription machinery, thereby increasing the steady state levels for fgl2 transcripts. an  
10 alternative mechanism may be N protein binding directly to fgl2 mrna, thus altering the rate of fgl2 transcript degradation.

The inventors have now shown that the mechanism by which N protein induces fgl2 prothrombinase expression most likely includes the phosphorylation of LF-A1, transport of the LF-A1 protein into the nucleus of  
15 infected cells, acting to modulate the signal transduction pathways that regulate host transcription machinery, thereby increasing the steady state levels for fgl2 transcripts. Recent work by both nuclear runoff assays and transient transfection experiments have demonstrated that the induction of fgl2 mRNA by MHV-3 infection is at least in part attributable to new  
20 transcription (Leibowitz, J.L. et al., in press).

Preliminary mapping of the fgl2 promoter has defined a region from -372 to -306 to be responsive to the N protein. The present inventors identified three putative cis-elements and have shown that LF-A1 binding element is the CIS element necessary for FGL-2 transcription. It was also  
25 shown that lf-ai is present in macrophages. Previous reports which have suggested that LF-A1 was typical of a liver specific gene and liver-enriched transcription factor. Furthermore, LF-A1 is known to regulate gene expression of coagulation factors IX, X and XII (Crossley et al., 1992; Citarella et al., 1993; Miao et al., 1992). The present data shows that GMCSF is not  
30 involved in inducing fgl-2, however, IE1.2 may play a minor role. IE1.2 has

been implicated as human cytomegalovirus immediate early gene 1 and 2 regulatory elements, providing a link between virus infection and inflammation (Fickenscher et al., 1989).

Recent studies have shown that the severity of hepatic injury  
5 in patients with hepatitis B is related to the synthesis and expression of  
nucleocapsid protein of HBV, suggesting that the accumulation of hepatitis  
B core antigen may damage hepatocytes directly or may serve to stimulate  
cell mediated immune responses (Davies, S.E. et al., 1991; Chisar, F.V. and  
Ferrari, C., 1995). The inventor has recently cloned and sequenced the  
10 human prothrombinase gene (hfgl2) gene and have shown its expression in  
the liver of 3 patients with fulminant hepatic failure). It is expected that the  
hepatitis B core antigen induces transcription of hfgl2.

In conclusion, mapping of genetic determinants in parenteral  
and recombinant MHV strains demonstrates that the N protein of strains of  
15 MHV which induce FHF is responsible for enhanced transcription of the  
fgl-2 prothrombinase gene. These studies provide significant insights into  
the viral pathogenesis of human diseases such as hepatitis B and C in which  
core antigen or pathogenic factor (nucleocapsid protein) influences disease  
activity.

20 While the present invention has been described with reference  
to what are presently considered to be the preferred examples, it is to be  
understood that the invention is not limited to the disclosed examples. To  
the contrary, the invention is intended to cover various modifications and  
equivalent arrangements included within the spirit and scope of the  
25 appended claims.

All publications, patents and patent applications are herein  
incorporated by reference in their entirety to the same extent as if each  
individual publication, patent or patent application was specifically and  
individually indicated to be incorporated by reference in its entirety.



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**DETAILED FIGURE LEGENDS**

**Figure 1.** The schematic representation of oligonucleotide maps of the recombinant viruses (MHV-A59 X JHM) and (MHV-A59 X MHV-2). L: leader sequence. Pol: RNA dependent RNA polymerase gene. HN: hemagglutinin-esterase glycoprotein gene. S: spike protein. M: matrix glycoprotein gene. N: nucleocapsid protein gene. The data are from Parler, M.M. and P.S. Masters, 1990.

**Figure 2.** A schematic diagram of the organization of the fgl2 gene is shown in the upper panel. The lower panel shows the sequence of the 1.3 kb DNA flanking 5' end of fgl2. The sequence has been deposited into Genbank with accession number AF025817. The sequence shows the overlapping 400 bp at 3' end of this fragment and the 5' end of the published sequence by Koyama et. al (Koyama et al., 1987) with the accession number M15761 are consistent. The putative cis-elements responsive to N protein and initiating ATG for translation are indicated in bold and underlined.

**Figure 3.** Expression of Balb/cJ macrophage functional PCA induced by parental viruses and their recombinants derived from MHV-A59 X JHM (A) or MHV-A59 X MHV-2 (B).

Macrophages from Balb/cJ were infected with viruses at a multiplicity of infection (MOI) of 2.5 for 8-10 h and harvested for measurement of PCA activity. Values represent the mean  $\pm$  SD of three separate experiments done in triplicate. \* represents  $p < 0.01$  when compared with unstimulated macrophages.

**Figure 4.** Expression of Balb/cJ macrophage fgl2 transcripts induced by parental viruses and their recombinants by RT-PCR. 5  $\mu$ g of total cellular RNA extracted from infected macrophages was reverse transcribed and then PCR was performed using specific fgl2 primers as described in Materials and

Methods. Lane 1, macrophages + MHV-3; Lane 2, macrophages + MHV-A59; Lane 3, macrophages + MHV-JHM; Lane 4, macrophages + MHV-2; Lane 5, macrophages + MHV-RL1; Lane 6, macrophages + MHV-CA13; Lane 7, macrophages + MHV-CA43; Lane 8, macrophages + MHV-ML3; Lane 9, macrophages + MHV-ML9; Lane 10, macrophages + MHV-ML11. PCA of glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was set up as a technique control.

**Figure 5.** Expression of nucleocapsid (N) protein in transfected cells by Western blot analysis.  $2 \times 10^6$  of cell lysis post transfection were loaded in a 10% of SDS-PAGE gel and transferred to a nitrocellulose membrane. Membrane were then probed with antibodies as described in Materials and Methods. Lane 1, CHO + fgl2 promoter construct + A59 N gene construct; Lane 2, CHO + fgl2 promoter construct + MHV-2 N gene construct; Lane 3, CHO + fgl2 promoter construct + pCR 3.1 vector alone; Lane 4. 17 CL1 cells + MHV-3.

**Figure 6.** Effect of N protein on fgl2 promoter. 0.5  $\mu$ g of N gene construct were cotransfected with 0.5  $\mu$ g of fgl2 promoter construct in CHO cells for 40-44 h and cells were harvested and freeze-thawed 3 times for measurement of luciferase activity. Values represent the mean  $\pm$  SD of five separate experiments done in triplicate. \* represents  $p < 0.01$  when compared with cells cotransfected with pCR 3.1 vector.

**Figure 7. A.** Transient expression of luciferase activity by deletion constructs of the fgl2 promoter in response to MHV-A59 N protein in CHO cells. A series of the fgl2 promoter constructs containing varying lengths of the fgl2 promoter sequence were cotransfected with a MHV-A59 N gene construct into CHO cells. Relative luciferase activity is expressed in fold increase compared to CHO cells cotransfected with the fgl2 promoter

constructs with empty pCR3.1 vector. PGL2-basic vector was used as a negative control. All luciferase assays represent the mean  $\pm$  SD of six or more independent experiments. \* represents  $p < 0.01$  when compared with cells cotransfected with empty pCR 3.1 vector.

- 5 B. Schematic representation of the putative regulatory elements in the putative ( -372 to -306) *fgl2* promoter responsive to N protein. Also shown are the ATG translation initiation site and the location of the TATA box.

**Figure. 8. Expression of Balb/cJ macrophage functional PCA induced by MHV and it I mutant.** Macrophages from Balb/cJ were infected with MHV and MHV-A59 I mutant (Alb 110 and it isogenic wide type Alb 111) at a M.O.I. of 2d.5 for 8-10 hrs and harvested for measurement of PCA activity. Values represent the mean  $\pm$  S.D. of three separate experiments done in triplicate. \* represents  $p < 0.01$  compared with unstimulated macrophages.

**Figure. 9. Transient expression of luciferase activity by *mfg12* promoter in response to MHV-A59 N protein and it mutants in CHO cells.** N gene constructs from MHV-A59 and MHV-2 and a series of N gene mutants from MHV-A59 was cotransfected with the wide type *fgl2* promoter respectively into CHO cells. Relative luciferase activity is expressed in fold increase compared with CHO cells cotransfected with N construct from MHV-2. PGL2-basic vector was used as a negative control. Values represent the mean  $\pm$  S.D. of five separate experiments done in triplicate. \* represents  $p < 0.01$  compared with cells cotransfected with MHV-2 N construct. # represents  $p < 0.01$  compared with cells cotransfected with N gene construct from wide type MHV-A59.

25 **Figure. 10. Confocus Microscope Immunofluoresence for detection of N protein.** Macrophages from Balb/cJ mice were planted as a monolayer on the glass slide flask



(VWR) and infected with MHV-A59 and MHV-2. Cells were stained for nucleocapsid protein in the nucleus of infected macrophages.

**Figure. 11. Electrophoresis mobility shift assays (EMSA) and competition studies.** Band shift assays were performed with oligonucleotides corresponding to the LF-A1 (lanes 1-4) or IE1.2 (lanes 5-8) or GMCSF (lanes 9-12) binding sites within the promoter region of fg12 gene. For competition experiments, nuclear extracts (Nu) were preincubated with either specific competitor (cold probes) or non specific competitor. After addition of radiolabel led probes, the free DNA and the DNA protein complex were separated on a 6% polyacrylamide gel. The sequences of the oligonucleotides used are shown in the description. Lane 1,5,9: Free 32P-probe; lane 2,6,10: 32P-probe + Nu; lane 3,8,11: 32P-probe + cold specific competitor + Nu; lane 4,7,12: 32P-probe + non specific competitor + Nu.

**Figure. 12. Transient expression of luciferase activity by mfg12 promoter and its mutants for candidate cis-elements in response to MHV-A59 N protein in CHO cells.** 0.5 ug of N gene constructs from MHV-A59, MHV-2 was cotransfected with 0.25 ug of wildtype (WT) pfg12 (-1328) LUC or its mutants for candidate cis elements LF-A1 (LFA1mut), IE1.2 (IEI1.2mut) or LFA1 and IE1.2 double mutation (LFA1/IE1.2mut) respectively in CHO cells for 40-44 hrs, cells were harvested and freeze-thawed three time for measurement of luciferase activity. PGL2-basic vector was used as a negative control. Values represent the mean  $\pm$  S.D. of four separate experiments done in triplicate. \* represents  $p < 0.01$  compared with cells cotransfected with MHV-2 N construct. # represents  $p < 0.01$  compared with cells cotransfected with wide type pfg12 (-1328) LUC.

**Figure. 13. Western blot analysis for detection of LF-A1 protein in macrophages in Balb/cJ.** Nuclear proteins (Nu) were extracted from

macrophages infected with MHV at a multiplicity of infection (M.O.I) of 2.5. CHO cells transfected with HNF-4 expression construct were used as positive control. Twenty (20)  $\mu$ l of lysate were resolved by SDS-PAGE and then transferred to a nitrocellulose membrane. Membrane was probed with a polyclonal antibody against the HNF-4 protein at 4°C overnight, followed by incubation with goat anti rabbit IgG labeled with horseradish peroxidase as described in the Materials and Methods of the description. Lane 1. HNF-4 expression construct + CHO Nu; lane 2. M $\phi$  Nu; lane 3. M $\phi$  + MHV-A59 Nu; lane 4. M $\phi$  + MHV-2 Nu.

**TABLE 1****Identity comparison of N gene sequence**

	M35156 (1.666 Kb)	MHV-2 (1.657 Kb)
MHV-A59 (1.666Kb)	99%	93%
MHV-ML3 (1.666 Kb)	99%	92%
MHV-ML11 (1.657Kb)	93%	99%

Multiple clones from each construct were sequenced. T7 primer and pCR3.1 reverse primer were used for 5' to 3' and 3' to 5' sequence, respectively. A new primer at position 557 was also designed to accomplish the cloned entire N gene sequence as described in Materials and Methods. The sequence was analyzed using the DNAsis for windows, sequence analysis software.

WE CLAIM:

1. A method of preventing or reducing immune coagulation caused by a virus comprising administering an effective amount of an inhibitor of a nucleocapsid gene (N-gene) or protein (N-protein) to an  
5 animal in need thereof.
2. A method according to claim 1 wherein the virus is a hepatitis virus.
3. A method according to claim 2 wherein the hepatitis virus is capable of inducing fgl-2.
- 10 4. A method according to claim 1 wherein the inhibitor of the N-protein is an antibody that binds to the N-protein or domain I of the N-protein.
5. A method according to claim 1 wherein the inhibitor of the N-gene is an antisense oligonucleotide that is complimentary to a nucleic acid  
15 sequence from the N-gene.
6. A method of inducing immune coagulation comprising administering an effective amount of a N-protein, domain I of the N-protein, or gene to an animal in need thereof.
7. A composition for use in inhibiting procoagulant activity in an  
20 animal comprising (a) an antibody specific for a N-protein or domain I of the N-protein; (b) or antisense nucleic acid molecules complimentary to the N-protein gene.

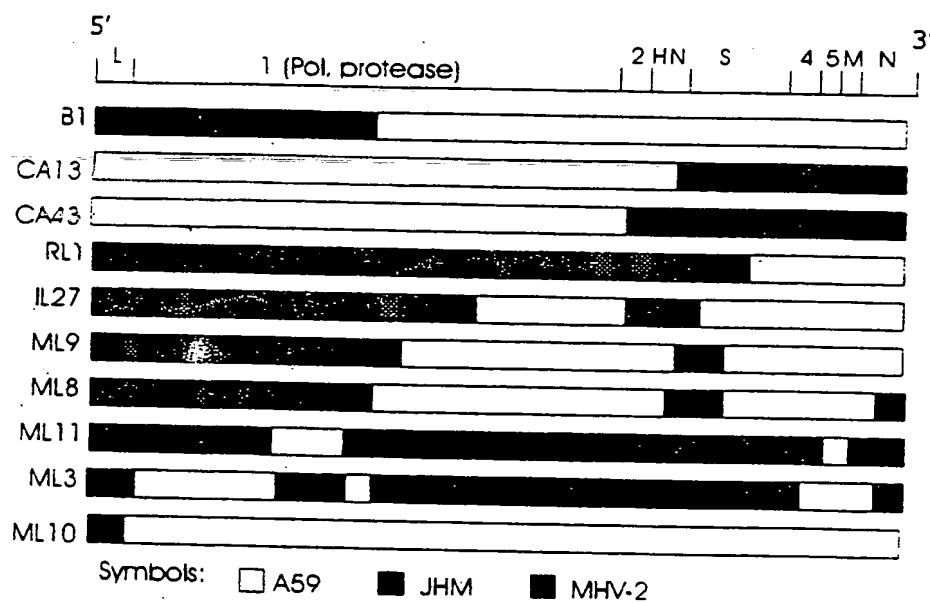
8. A composition for use in inducing procoagulant activity in an animal comprising a nucleic acid sequence encoding N-protein or domain I of N-protein; or an N-protein or domain 1 of N-protein in admixture with a suitable diluent or carrier.
- 5 9. A vaccine against a virus causing immune coagulation comprising an amount of an N-protein or peptide which is effective to induce an immune response against N-protein.
10. A vaccine for treating or preventing viral hepatitis comprising an effective amount of an N-protein or peptide in admixture with a suitable  
10 diluent or carrier.
11. A method of preventing or reducing immune coagulation caused by a virus comprising administering an effective amount of an inhibitor of LF-A1 gene or protein to an animal in need thereof.
12. A method according to claim 11 wherein the virus is a hepatitis  
15 virus.
13. A method according to claim 11 wherein the inhibitor of the LF-A1 protein is an antibody that binds to the LF-A1 protein.
14. A method according to claim 1 wherein the inhibitor of LF-A1 protein is an antisense oligonucleotide that is complimentary to the LF-A1  
20 binding element of the promoter region of the fgl-2 gene.
15. A method according to claim 11 wherein the inhibitor of the LF-A1 gene is an antisense oligonucleotide that is complimentary to a nucleic acid sequence from the LF-A1 gene.

16. A method of inducing immune coagulation comprising administering an effective amount of a LF-A1 protein or gene to an animal in need thereof.

17. A composition for use in inhibiting procoagulant activity in an  
5 animal comprising (a) an antibody specific for a LF-A1 protein; (b) or antisense nucleic acid molecules complimentary to the LF-A1 protein gene; or (c) antisense nucleic acid molecules complimentary to the LF-A1 binding element of the promoter region of the fgl-2 gene.

18. A composition for use in inducing procoagulant activity in an  
10 animal comprising a nucleic acid sequence encoding LF-A1 protein or an LF-A1 protein in admixture with a suitable diluent or carrier.

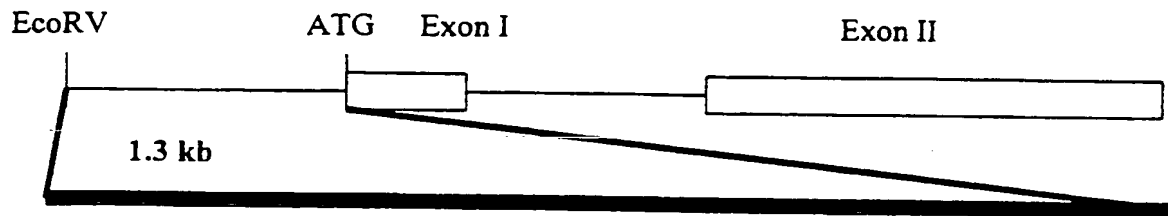
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**FIGURE 1**

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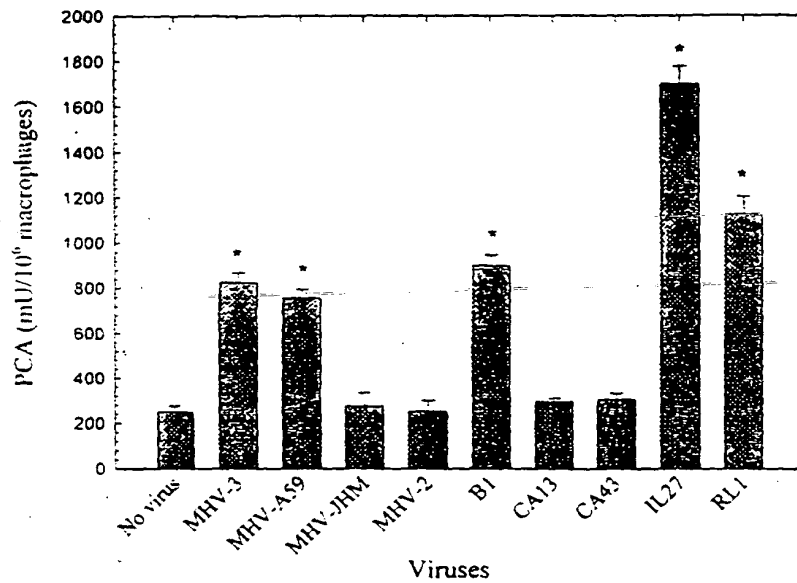
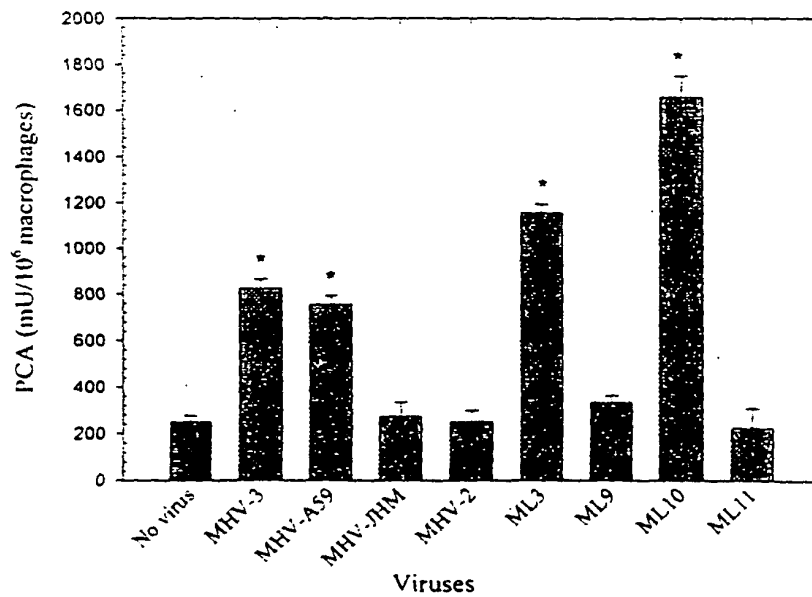
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**FIGURE 2**

GATATCATGG	GATGGAATGA	GAAGGGAAAG	TAGGAGCCCC	AGAGTGCGGT	AAGACAAGGC	60
ATAAGGCGTG	TCTGACAAAT	TCTTCATACA	CACATTTCCC	CTTTGCACAT	TCAGTCTGTA	120
TAGGTTATTT	CTATAGGAGA	AAAAAATAT	TCAAATTCCT	TGTGCACTGG	TAACAGGCAT	180
GAAGGCTCAG	CAAAGCCAAT	ACGTGTTATG	TCCAGTTGGA	GACAGTGCCA	GGGCCAACAT	240
TCCAGACTTC	TCAGATAGAA	AGTGCGCCTG	CCTGCCCTGC	TCTGAGAATT	TGAAGAGAGT	300
AGTTCAGTTA	GAATTAAGAG	GCAGTAGAGA	AAAGTCTTGG	GAAATCTGGT	TAGAGATATA	360
AATATGAGAA	CTGGACATGG	TGGTACACAC	CTGTGATCTC	TGTGTTTAGG	AGGGAGAGGC	420
AGAGAGATCA	GGAGTTCAAG	GCCAGCCTGA	GCTACTTGAG	ACCCAGTCTA	AATAAATAAG	480
AGATAGATTA	CAGAGTGCCT	TTAACTAGTA	CAGAGAAAGA	ATTTGGGTTT	ATCTGTGTCA	540
GTTACGCTGA	AATAATTTTT	AAGTAATAAA	ATCCCTTTTA	ATAAGAAACC	TTATGAGGTC	600
AGTATGCACA	ATGAACCTAA	GAGAGACCCC	CAGCTCCTGA	GCTGAGTGAT	GGGGAAGGAC	660
AGCCACTGCC	TGTGATGTGT	GAGTGACGTG	CTTCCAAGTG	TTTAAACCAC	TGACGATTAC	720
ATAGCCTGCA	CAGTCAGGAG	AAAACAGCCG	TATTCTCTGC	CAGTTCTCTT	CCCTTTTACA	780
AACAGATGAG	AGACACACAC	AGAGAATCCA	TTTAAAGAGC	GGACCTTTGT	TCTGATTAGG	840
GGCAATTTTA	AGTACTTAAG	AGTTCACACA	AAGTCTAGCC	TTCAAAAAGA	AAACAGGTTC	900
CCAAACTAGG	GAGGAAACAG	AATCATTTC	ATTTTGGTGA	CATTTAGTGG	GAAGAAGCTC	960
ACAGACATTT	AGACGTTCCA	ACTCTTTCCC	CAGTAGTGGA	CCAAGTATAT	AATATGGTAT	1020
GMCSF		IE1.2	LF-A1			
CTTTTGGGCA	CTGGTATTAC	AACTGTTTTT	TAAACAAAAG	ACTTTCCTTG	TGCTTTACTA	1080
AAAACCCAGA	CGGTGAATCT	TGAATACAAT	GCGTGGCACC	CACGGCAGGC	ATTCTATTGT	1140
GCATAGTTTT	GAATGACAGG	AGATGACAGC	ATTTGGCTGG	CTGCGCTTGC	TGAGGACCCT	1200
CTCCTCCTGT	GTGGCGTCTG	AGACTGTGAT	GCAAATGCGC	CCGCCCTTTT	CTGGGAAGTC	1260
		SP-1				
AGAACGCCTG	AGTCAGGCGG	CGGTGGCTAT	TAAAGCGCCT	GGTCAGGCTG	GGCTGCCGCA	1320
AP-1		TATA-box				
CTGCAAGGATG						

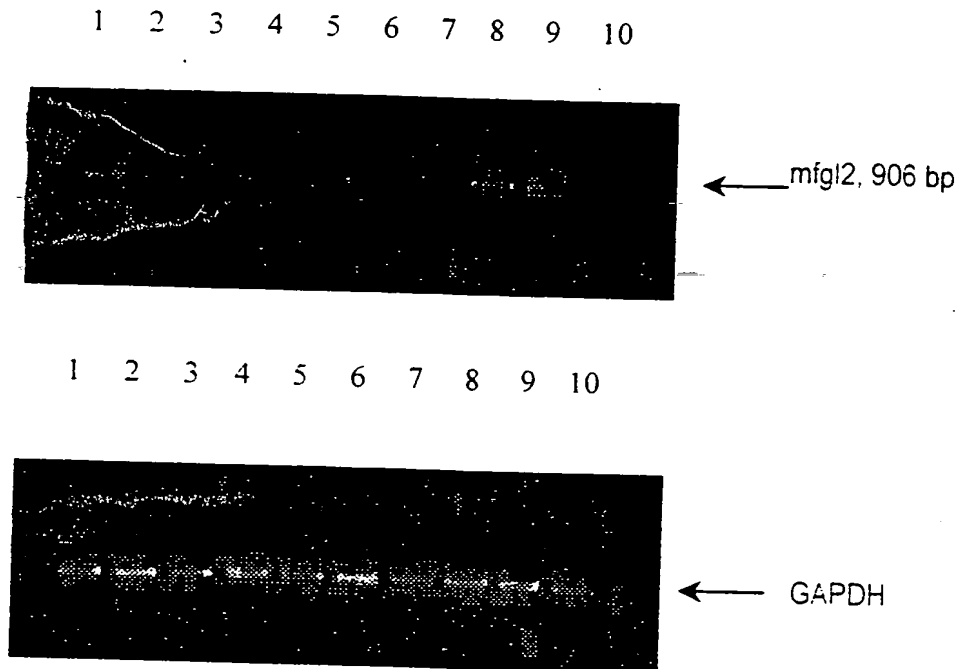
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**FIGURE 3****A****B**

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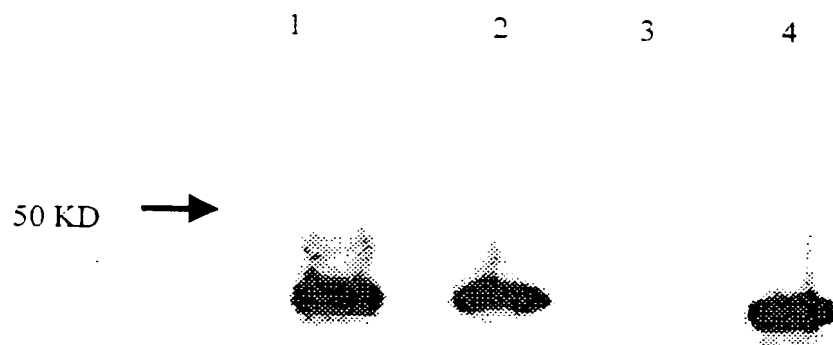
4/13

**FIGURE 4**

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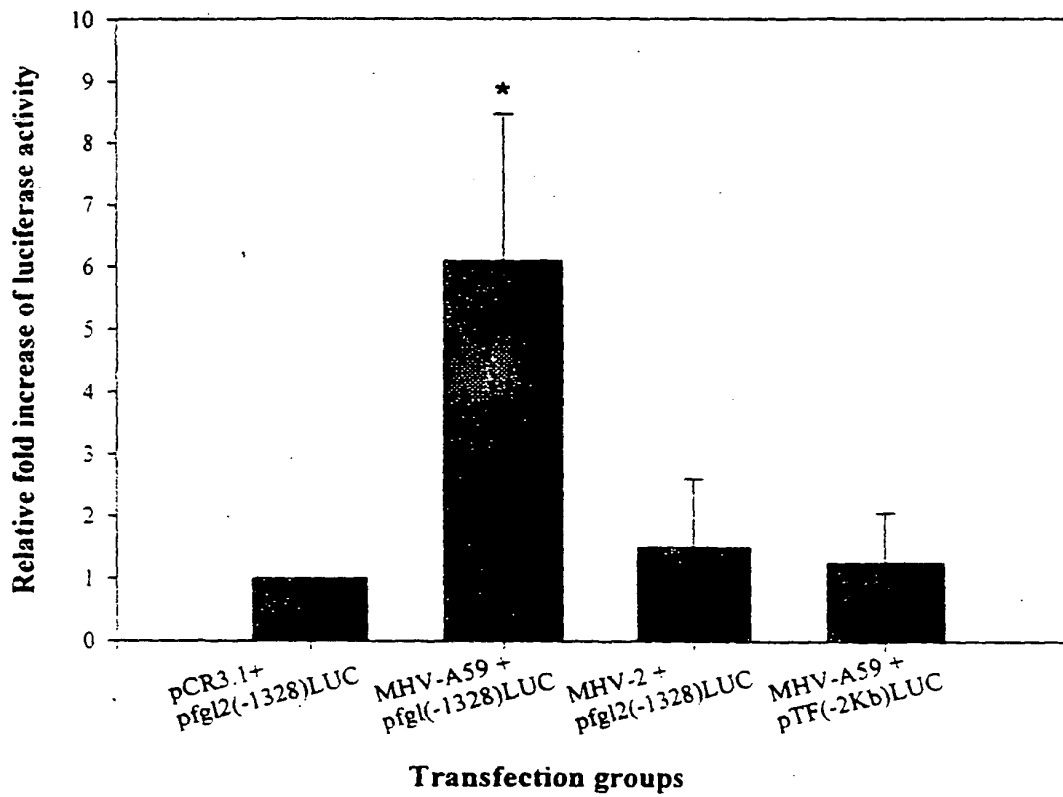
**FIGURE 5**



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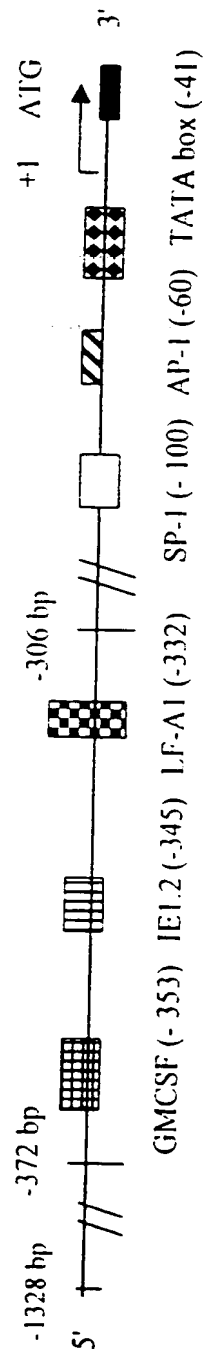


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**FIGURE 6**

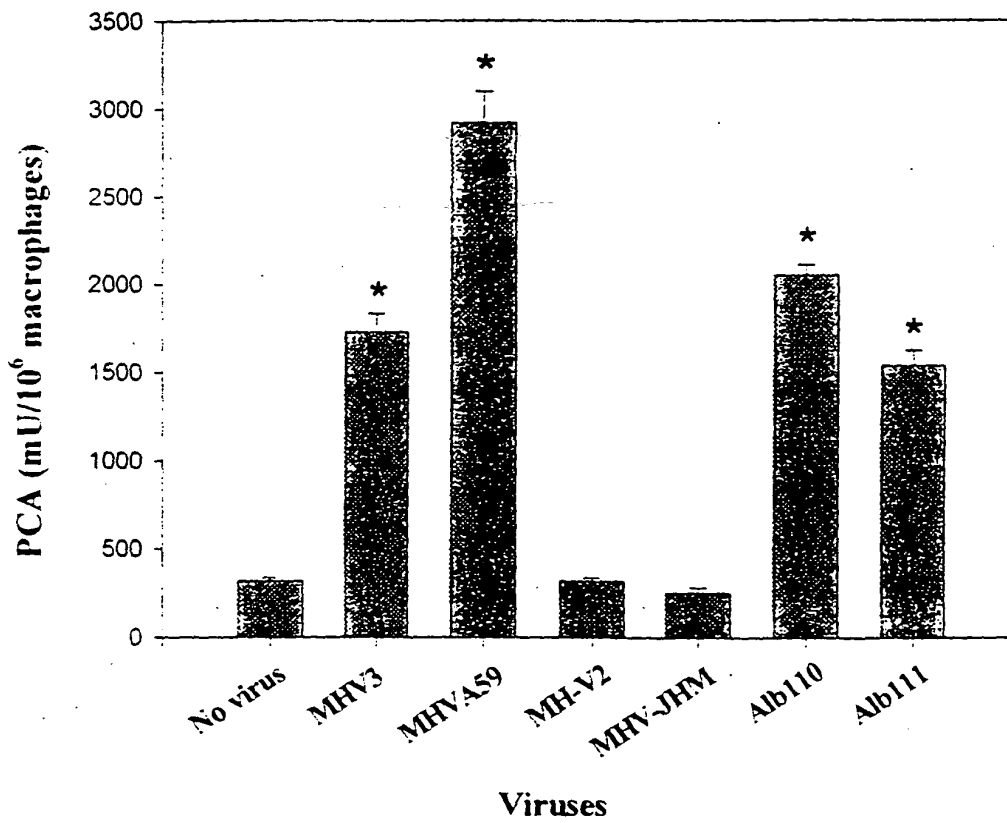
518 Rec'd PCT/PTO 24 AUG 2001

### FIGURE 7



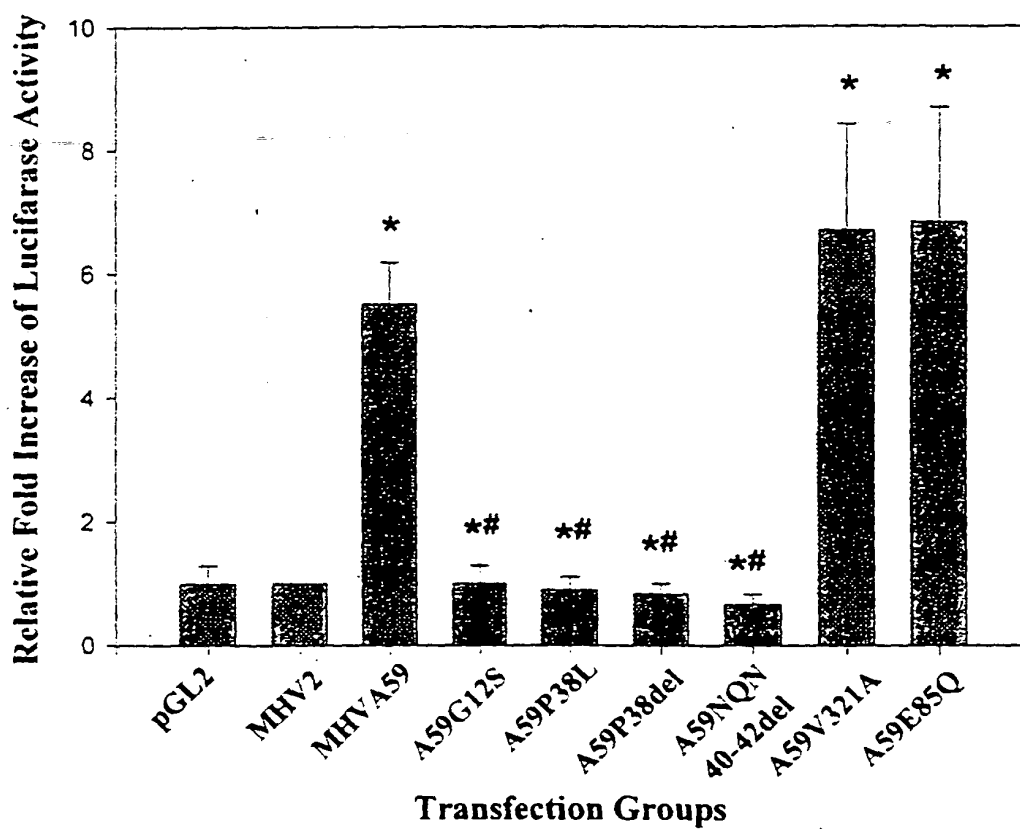
518 Rec'd PCT/PTO 24 AUG 2001

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**FIGURE 8**

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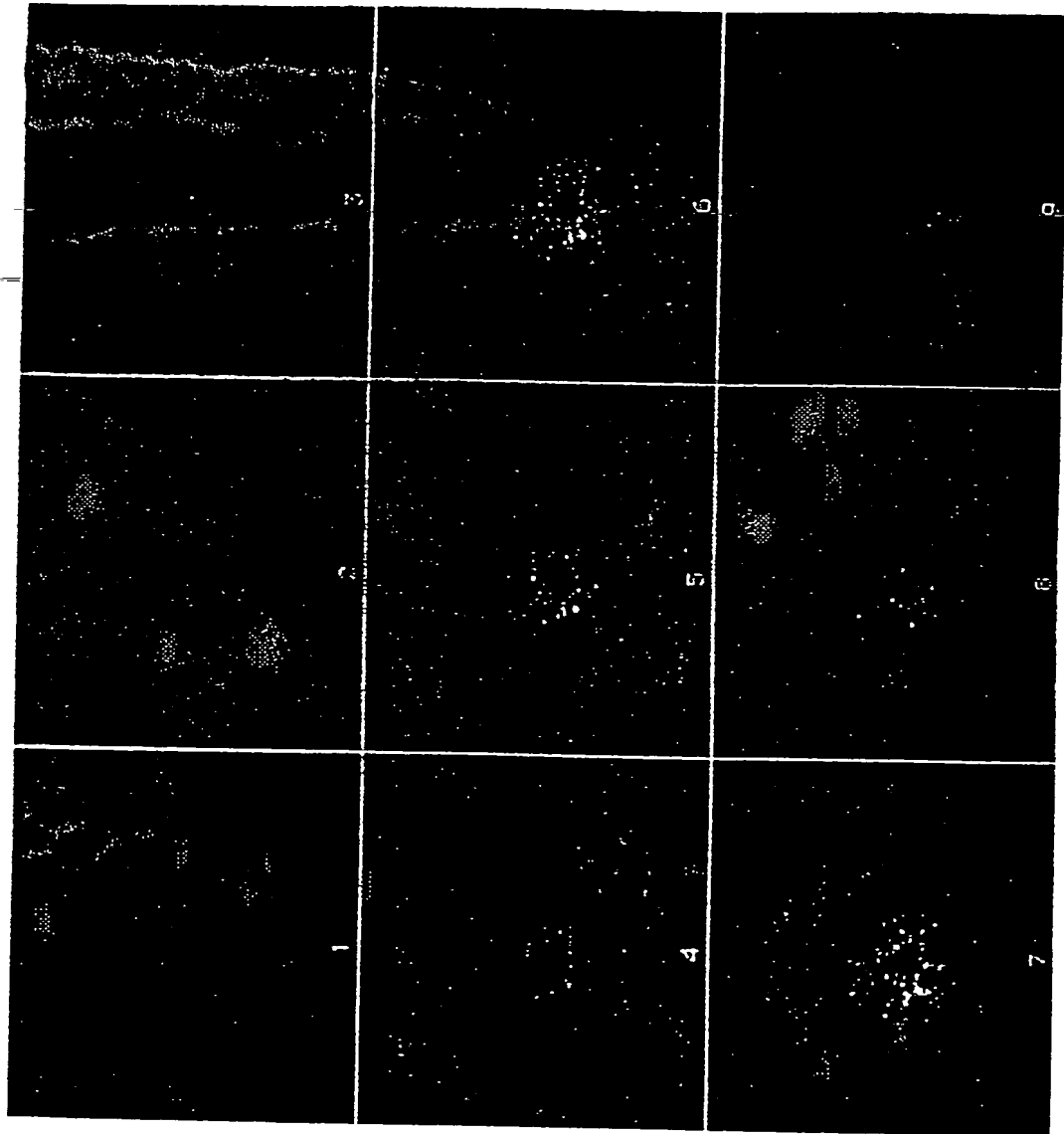
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**FIGURE 9**

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FIGURE 10

057

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**FIGURE 11**

1 2

3 4

5

6

7

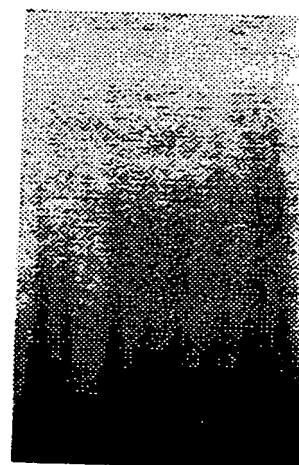
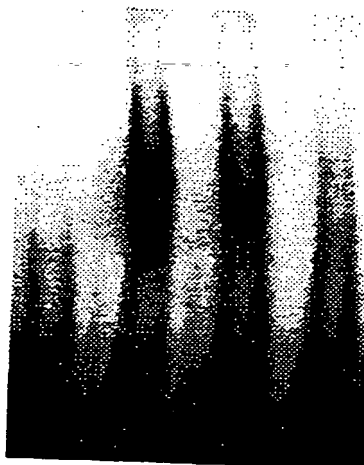
8

9

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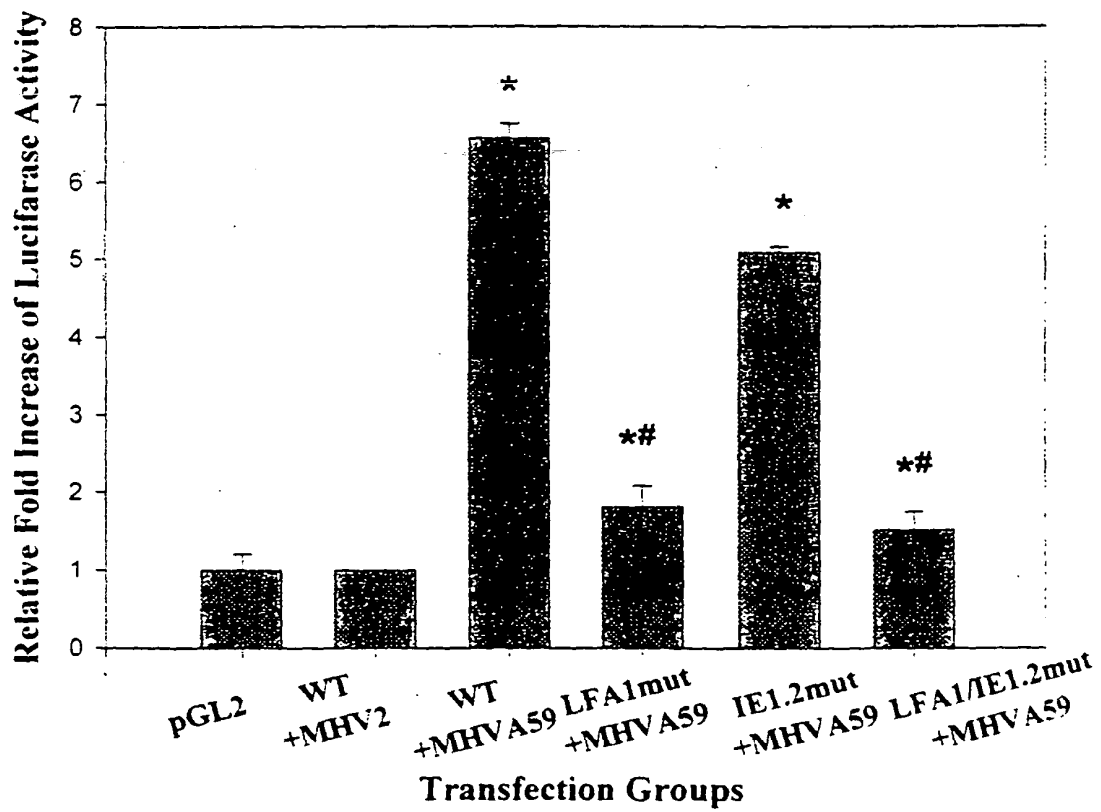
11

12



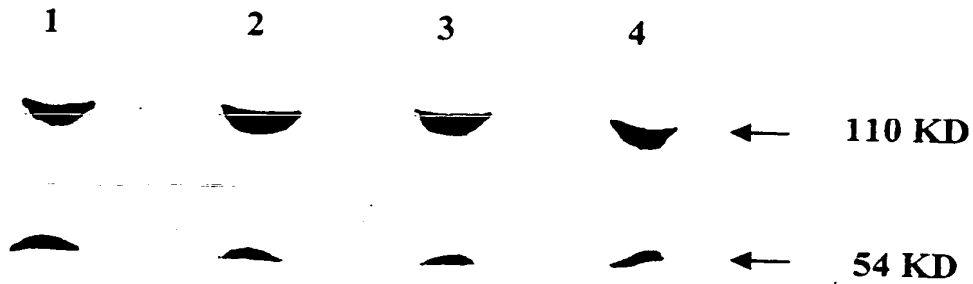
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**FIGURE 12**

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**FIGURE 13**

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## INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

Applicant's or agent's file reference 9579-24	<b>FOR FURTHER ACTION</b> See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)	
International application No. PCT/CA00/00191	International filing date (day/month/year) 25/02/2000	Priority date (day/month/year) 26/02/1999
International Patent Classification (IPC) or national classification and IPC A61K39/395		
Applicant LEVY, Gary		

1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.



2. This REPORT consists of a total of 8 sheets, including this cover sheet.

- ☐ This report is also accompanied by ANNEXES, i.e. sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).

These annexes consist of a total of sheets.

3. This report contains indications relating to the following items:

- I ☒ Basis of the report
- II ☐ Priority
- III ☒ Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
- IV ☐ Lack of unity of invention
- V ☒ Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement
- VI ☐ Certain documents cited
- VII ☒ Certain defects in the international application
- VIII ☒ Certain observations on the international application

Date of submission of the demand 18/09/2000	Date of completion of this report 09.05.2001
Name and mailing address of the international preliminary examining authority:  European Patent Office D-80298 Munich Tel. +49 89 2399 - 0 Tx: 523656 epmu d Fax: +49 89 2399 - 4465	Authorized officer BROCHADO GARGANTA, M Telephone No. +49 89 2399 8935 



# INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No. PCT/CA00/00191

## I. Basis of the report

1. With regard to the **elements** of the international application (*Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to this report since they do not contain amendments (Rules 70.16 and 70.17)*):

**Description, pages:**

1-45 as originally filed

**Claims, No.:**

1-18 as originally filed

**Drawings, sheets:**

1/13-13/13 as originally filed

**Sequence listing part of the description, pages:**

1-4, filed with the letter of 13.07.2001

2. With regard to the **language**, all the elements marked above were available or furnished to this Authority in the language in which the international application was filed, unless otherwise indicated under this item.

These elements were available or furnished to this Authority in the following language: , which is:

- ☐ the language of a translation furnished for the purposes of the international search (under Rule 23.1(b)).
- ☐ the language of publication of the international application (under Rule 48.3(b)).
- ☐ the language of a translation furnished for the purposes of international preliminary examination (under Rule 55.2 and/or 55.3).

3. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international preliminary examination was carried out on the basis of the sequence listing:

- ☐ contained in the international application in written form.
- ☐ filed together with the international application in computer readable form.
- ☒ furnished subsequently to this Authority in written form.
- ☒ furnished subsequently to this Authority in computer readable form.
- ☒ The statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.
- ☒ The statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished.

4. The amendments have resulted in the cancellation of:



# INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No. PCT/CA00/00191

- ☐ the description, pages:
- ☐ the claims, Nos.:
- ☐ the drawings, sheets:

5. ☐ This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)):

*(Any replacement sheet containing such amendments must be referred to under item 1 and annexed to this report.)*

6. Additional observations, if necessary:

### III. Non-establishment of opinion with regard to novelty, inventive step and industrial applicability

1. The questions whether the claimed invention appears to be novel, to involve an inventive step (to be non-obvious), or to be industrially applicable have not been examined in respect of:

☐ the entire international application.

☒ claims Nos. 1-6, 11-16.

because:

☒ the said international application, or the said claims Nos. 1-6, 11-16, with respect to industrial applicability relate to the following subject matter which does not require an international preliminary examination (*specify*):

**see separate sheet**

☐ the description, claims or drawings (*indicate particular elements below*) or said claims Nos. are so unclear that no meaningful opinion could be formed (*specify*):

☐ the claims, or said claims Nos. are so inadequately supported by the description that no meaningful opinion could be formed.

☐ no international search report has been established for the said claims Nos. .

2. A meaningful international preliminary examination cannot be carried out due to the failure of the nucleotide and/or amino acid sequence listing to comply with the standard provided for in Annex C of the Administrative Instructions:

☐ the written form has not been furnished or does not comply with the standard.

☐ the computer readable form has not been furnished or does not comply with the standard.

### V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement



# INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No. PCT/CA00/00191

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## 1. Statement

Novelty (N)	Yes:	Claims	3, 4, 9-18
	No:	Claims	1, 2, 5-8
Inventive step (IS)	Yes:	Claims	
	No:	Claims	1-18
Industrial applicability (IA)	Yes:	Claims	7-10, 17-18
	No:	Claims	

## 2. Citations and explanations

**see separate sheet**

## VII. Certain defects in the international application

The following defects in the form or contents of the international application have been noted:  
**see separate sheet**

## VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:  
**see separate sheet**





## INTERNATIONAL SEARCH REPORT

Int. l. Application No.

PCT/CA 00/00191

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 A61K39/395 A61K38/16 C12N15/11 A61K48/00 A61K39/12  
 A61K39/29 A61P7/00 A61P7/02 //C07K16/28

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

MEDLINE, BIOSIS, WPI Data, EPO-Internal, PAJ

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>NING Q ET AL: "The nucleocapsid (N) protein of murine hepatitis virus type 3 (MHV-3) induces transcription of the novel fgl-2 prothrombinase gene."</p> <p>HEPATOLOGY, vol. 28, no. 4 PART 2, October 1998 (1998-10), page 405A XP000929770</p> <p>Biennial Scientific Meeting of the International Association for the Study of the Liver and the 49th Annual Meeting and Postgraduate Courses of the American Association for the Study of Liver Diseases; Chicago, Illinois, USA; November 4-10, 1998 ISSN: 0270-9139 the whole document</p> <p style="text-align: center;">---</p> <p style="text-align: center;">-/--</p>	1-10

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

## \* Special categories of cited documents:

\*A\* document defining the general state of the art which is not considered to be of particular relevance

\*E\* earlier document but published on or after the international filing date

\*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

\*O\* document referring to an oral disclosure, use, exhibition or other means

\*P\* document published prior to the international filing date but later than the priority date claimed

\*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

\*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

\*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

\*Z\* document member of the same patent family

Date of the actual completion of the international search

7 August 2000

Date of mailing of the international search report

23/08/2000

Name and mailing address of the ISA

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 NL - 2280 HV Rijswijk  
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Authorized officer

Covone, M

## INTERNATIONAL SEARCH REPORT

Int. lional Application No

PCT/CA 00/00191

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>STOHLMAN S A ET AL: "Location of antibody epitopes within the mouse hepatitis virus nucleocapsid protein."            VIROLOGY, (1994 JUL) 202 (1) 146-53. ,            XP002144399            abstract            page 151, right-hand column, line 15 -page            152, left-hand column, line 18</p>	7-10
X	<p>DATABASE MEDLINE 'Online!            US NATIONAL LIBRARY OF MEDICINE (NLM),            BETHESDA, MD, US;            MIZUTANI T ET AL: "Both antisense and            sense RNAs against the nucleocapsid            protein gene inhibit the multiplication of            mouse hepatitis virus."            retrieved from STN            Database accession no. 94355440            XP002144401            abstract            &amp; JOURNAL OF VETERINARY MEDICAL SCIENCE,            (1994 APR) 56 (2) 211-5. ,</p>	7,8
X	<p>US 5 849 485 A (ZHONG WEIMIN ET AL)            15 December 1998 (1998-12-15)            column 1, line 25-39            column 30, line 56 -column 31, line 4</p>	17,18
A	<p>DING J W ET AL: "Cloning and            characterization of the human            prothrombinase gene (H-fgl2) and its role            in human hepatitis."            HEPATOLOGY,            vol. 28, no. 4 PART 2,            October 1998 (1998-10), page 254A            XP000929678            Biennial Scientific Meeting of the            International Association for the Study of            the Liver and the 49th Annual Meeting and            Postgraduate Courses of the American            Association for the Study of Liver            Diseases; Chicago, Illinois, USA; November            4-10, 1998            ISSN: 0270-9139            the whole document</p>	1-18

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# INTERNATIONAL SEARCH REPORT

Int'l Application No  
PCT/CA 00/00191

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	<p>NING QIN ET AL: "The nucleocapsid protein of murine hepatitis virus type 3 induces transcription of the novel fgl2 prothrombinase gene." JOURNAL OF BIOLOGICAL CHEMISTRY. vol. 274, no. 15, 9 April 1999 (1999-04-09), pages 9930-9936, XP002144400 ISSN: 0021-9258 the whole document</p>	1-18
P,X	<p>LEVY GARY A ET AL: "Study of putative CIS-elements in MFGL2 promoter in response to nucleocapsid protein of murine hepatitis virus type 3." HEPATOLOGY, vol. 30, no. 4 PART 2, October 1999 (1999-10), page 502A XP000929772 50th Annual Meeting and Postgraduate Courses of the American Association for the Study of Liver Diseases; Dallas, Texas, USA; November 5-9, 1999 ISSN: 0270-9139 the whole document</p>	1-18

# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/CA 00/00191

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US 5849485 A	15-12-1998	US 5604115 A	18-02-1997
		US 6025196 A	15-02-2000
		AT 185598 T	15-10-1999
		AU 665939 B	25-01-1996
		AU 9174291 A	22-07-1992
		CA 2098838 A	22-06-1992
		DE 69131718 D	18-11-1999
		DE 69131718 T	02-03-2000
		EP 0564592 A	13-10-1993
		JP 6505152 T	16-06-1994
		WO 9211365 A	09-07-1992

**Re Item I**

**Basis of the report**

The Sequence listing pages 1-4 are not considered to be part of the application as they were not filed with the application (Rule 13ter.1-(f) PCT).

**Re Item III**

**Non-establishment of opinion with regard to novelty, inventive step and industrial applicability**

Claims 1-6 and 11-16 are directed to a method of treatment of the human/animal body. They relate therefore to subject-matter considered by this Authority to be covered by the provisions of Rule 67.1(iv) PCT. Consequently, no opinion will be formulated with respect to the industrial applicability of the subject-matter of these claims (Article 34(4)(a)(i) PCT).

**Re Item V**

**Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement**

1. Reference is made to the following documents:

- (B) Ning Q et al.: Hepatology, vol. 28, no. 4 Part 2, October 1998 (1998-10), page 405A
- (C) Stohlman S. et al.: Virology 202, 146-153 (1994)
- (D) Ning Q et al.: Journal of Biological Chemistry, vol. 274, no. 15, 9 April 1999 (1999-04-09), pages 9930-9936
- (E) Levy Gary et al.: Hepatology, vol. 30, no. 4 Part 2, October 1999 (1999-10), page 502A



**INTERNATIONAL PRELIMINARY  
EXAMINATION REPORT - SEPARATE SHEET**

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International application No. PCT/CA00/00191

2. Documents D and E cited in the Search Report as intermediate documents are to be considered as state of the art for claims 11-18, as the date of priority is not validly claimed for these claims.

The administration of an inhibitor of LF-A1 gene or protein or a composition containing it, are not disclosed in the priority document.

3. Novelty

- 3.1 The subject-matter of claims 1, referring to a method for preventing or reducing immune coagulation caused by a virus, is not new in the sense of Article 33(2) PCT, because the method with these features is already disclosed in document A.

Document A discloses a method for inhibiting viral replication, wherein antisense nucleic acid molecules against the nucleocapsid protein gene of mouse hepatitis virus are administered (see abstract).

- 3.2 The same applies to dependent claims 2 and 5. Their features are already known from document A (see abstract).

- 3.3 The subject-matter of claims 6, referring to a method for inducing immune coagulation, is not new in the sense of Article 33(2) PCT, because the method with these features is already known from document B.

Document B discloses a method for inducing immune coagulation, wherein the nucleocapsid (N) protein of murine hepatitis virus type 3 induces transcription of the fgl-2 prothrombinase gene, resulting in an increased functional procoagulant activity (see abstract).

- 3.4 Claims 7 and 8, relate respectively to a composition for use in inhibiting and inducing procoagulant activity. Such compositions are already known from documents A and B (see abstract).

4. Inventive step





**INTERNATIONAL PRELIMINARY  
EXAMINATION REPORT - SEPARATE SHEET**

---

International application No. PCT/CA00/00191

- 4.1 Claims 3 and 4 do not contain any additional features which in combination with the features of claim 1, meet the requirements of the PCT in respect of inventive step (Article 33(3) PCT).

The additional features of these claims, i.e. the induction of fgl-2 by the hepatitis virus and having an antibody that binds to the N-protein or to domain I of the N-protein, are well known in the art. The induction of fgl-2 is an functional characteristic inherent of the nucleocapsid protein of hepatitis virus (see for example document B, title). The use of antibodies as inhibitor of protein N will be obvious for the skilled person, as they would have a high affinity for this protein. They are also normally used in the art (see for example document C, abstract).

- 4.2 Claims 9 and 10 relate to a vaccine comprising an effective amount of an N-protein. It is known in the art that hepatitis virus N protein, which forms the viral helical nucleocapsids, performs a number of regulatory functions during virus replication (see for example the abstracts of documents A, B and C). Thus, it would be obvious for the skilled person to use this protein for a vaccine against hepatitis. Thus, claims 9 and 10 are not based on an inventive concept as required by Article 33(3) PCT.

- 4.3 Claim 11 refers to a method for preventing or reducing immune coagulation, wherein an effective amount of an inhibitor of LF-A1 gene or protein is administered.

It is known from document D that there are three putative cis-elements in the fgl2 promoter, that are necessary for transcription. One of these elements is the LF-A1, which is known to regulate gene expression of coagulation factors (see page 9934, column 2). Thus, the skilled person would consider the use of an inhibitor of this element for the method of claim 1, without for that the need of an inventive concept. Therefore, claim 11 is not inventive (Article 33(3) PCT).

- 4.4 Considering the reasoning given in 4.3 and with regard to the reasoning given in this communication for claims 1-10, it is considered that claims 12-18 are also not based on an inventive step as required by Article 33(3) PCT.

5. For the assessment of the present claims 1-6 and 11-16 on the question whether they are industrially applicable, no unified criteria exist in the PCT Contracting States.



**INTERNATIONAL PRELIMINARY  
EXAMINATION REPORT - SEPARATE SHEET**

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International application No. PCT/CA00/00191

The patentability can also be dependent upon the formulation of the claims. The EPO, for example, does not recognize as industrially applicable the subject-matter of claims to the use of a compound in medical treatment, but may allow, however, claims to a known compound for first use in medical treatment and the use of such a compound for the manufacture of a medicament for a new medical treatment.

**Re Item VII**

**Certain defects in the international application**

1. Dependent claim 14 has a back reference to claim 1, but it is more probably depending on claim 11.

**Re Item VIII**

**Certain observations on the international application**

1. The use of general statements in the description which imply that the extent of protection may be expanded in some vague and not precisely defined way ("spirit of the invention" on pages 4, line 20, and 34, line 24), is not allowable (Article 6 PCT, see Guidelines III-4.3.a).





## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification 7 :</b> A61K 39/395, 38/16, C12N 15/11, A61K 48/00, 39/12, 39/29, A61P 7/00, 7/02 // C07K 16/28	<b>A1</b>	<b>(11) International Publication Number:</b> WO 00/51636 <b>(43) International Publication Date:</b> 8 September 2000 (08.09.00)
<b>(21) International Application Number:</b> PCT/CA00/00191 <b>(22) International Filing Date:</b> 25 February 2000 (25.02.00) <b>(30) Priority Data:</b> 60/122,109 26 February 1999 (26.02.99) US <b>(71)(72) Applicant and Inventor:</b> LEVY, Gary [CA/CA]; 260 Henderson Avenue, Thornhill, Ontario L3T 2M2 (CA). <b>(74) Agent:</b> BERESKIN & PARR; 40th floor, 40 King Street West, Toronto, Ontario M5H 3Y2 (CA).		<b>(81) Designated States:</b> AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).  <b>Published</b> <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
<b>(54) Title:</b> MODULATORS OF FGL2 PROTHROMBINASE  <b>(57) Abstract</b>  The inventor has determined that the nucleocapsid protein from a hepatitis virus can induce the prothrombinase, fgl2. The inventor has further developed that LF-A1 also induces fgl-2. This allows for the development of therapeutic methods and compositions for modulating immune coagulation. In particular, inhibitors of the N-protein or gene or LF-A1 protein gene, or LF-A1 binding site on the fgl-2 promoter may be useful in inhibiting immune coagulation caused by a virus such as a hepatitis virus.		

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Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

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PCT

NOTICE INFORMING THE APPLICANT OF THE  
COMMUNICATION OF THE INTERNATIONAL  
APPLICATION TO THE DESIGNATED OFFICES

(PCT Rule 47.1(c), first sentence)

From the INTERNATIONAL BUREAU

To:

BERESKIN & PARR  
40th floor  
40 King Street West  
Toronto, Ontario M5H 3Y2  
CANADA

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SEP 19 2000

BERESKIN &amp; PARR

SEP 20 2000

Date of mailing (day/month/year) 08 September 2000 (08.09.00)		IMPORTANT NOTICE	
Applicant's or agent's file reference 9579-24			
International application No. PCT/CA00/00191	International filing date (day/month/year) 25 February 2000 (25.02.00)	Priority date (day/month/year) 26 February 1999 (26.02.99)	
Applicant LEVY, Gary			

1. Notice is hereby given that the International Bureau has communicated, as provided in Article 20, the international application to the following designated Offices on the date indicated above as the date of mailing of this Notice:  
**AU, KP, KR, US**

In accordance with Rule 47.1(c), third sentence, those Offices will accept the present Notice as conclusive evidence that the communication of the international application has duly taken place on the date of mailing indicated above and no copy of the international application is required to be furnished by the applicant to the designated Office(s).

2. The following designated Offices have waived the requirement for such a communication at this time:  
**AE, AL, AM, AP, AT, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EA, EE, EP, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, OA, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW**  
The communication will be made to those Offices only upon their request. Furthermore, those Offices do not require the applicant to furnish a copy of the international application (Rule 49.1(a-bis)).
3. Enclosed with this Notice is a copy of the international application as published by the International Bureau on 08 September 2000 (08.09.00) under No. WO 00/51636

**REMINDER REGARDING CHAPTER II (Article 31(2)(a) and Rule 54.2)**

If the applicant wishes to postpone entry into the national phase until 30 months (or later in some Offices) from the priority date, a demand for international preliminary examination must be filed with the competent International Preliminary Examining Authority before the expiration of 19 months from the priority date.

It is the applicant's sole responsibility to monitor the 19-month time limit.

Note that only an applicant who is a national or resident of a PCT Contracting State which is bound by Chapter II has the right to file a demand for international preliminary examination.

**REMINDER REGARDING ENTRY INTO THE NATIONAL PHASE (Article 22 or 39(1))**

If the applicant wishes to proceed with the international application in the national phase, he must, within 20 months or 30 months, or later in some Offices, perform the acts referred to therein before each designated or elected Office.

For further important information on the time limits and acts to be performed for entering the national phase, see the Annex to Form PCT/IB/301 (Notification of Receipt of Record Copy) and Volume II of the PCT Applicant's Guide.

The International Bureau of WIPO 34, chemin des Colombettes 1211 Geneva 20, Switzerland	Authorized officer  J. Zahra
Facsimile No. (41-22) 740.14.35	Telephone No. (41-22) 338.83.38





**NOTICE INFORMING THE APPLICANT OF THE COMMUNICATION OF  
THE INTERNATIONAL APPLICATION TO THE DESIGNATED OFFICES**

<b>Date of mailing (day/month/year)</b> 08 September 2000 (08.09.00)	<b>IMPORTANT NOTICE</b>
<b>Applicant's or agent's file reference</b> 9579-24	<b>International application No.</b> PCT/CA00/00191

The applicant is hereby notified that, at the time of establishment of this Notice, the time limit under Rule 46.1 for making amendments under Article 19 has not yet expired and the International Bureau had received neither such amendments nor a declaration that the applicant does not wish to make amendments.



PCT

To:

BERESKIN & PARR  
40 King Street West  
40th Floor  
Toronto, Ontario M5H 3Y2  
CANADA

RECEIVED

MAY 16 2001

BERESKIN & PARR

NOTIFICATION OF TRANSMITTAL OF  
THE INTERNATIONAL PRELIMINARY  
EXAMINATION REPORT

(PCT Rule 71.1)

Date of mailing  
(day/month/year)

09.05.2001

Applicant's or agent's file reference  
9579-24

IMPORTANT NOTIFICATION

International application No.  
PCT/CA00/00191

International filing date (day/month/year)  
25/02/2000

Priority date (day/month/year)  
26/02/1999

Applicant

LEVY, Gary

1. The applicant is hereby notified that this International Preliminary Examining Authority transmits herewith the international preliminary examination report and its annexes, if any, established on the international application.
2. A copy of the report and its annexes, if any, is being transmitted to the International Bureau for communication to all the elected Offices.
3. Where required by any of the elected Offices, the International Bureau will prepare an English translation of the report (but not of any annexes) and will transmit such translation to those Offices.

4. REMINDER

The applicant must enter the national phase before each elected Office by performing certain acts (filing translations and paying national fees) within 30 months from the priority date (or later in some Offices) (Article 39(1)) (see also the reminder sent by the International Bureau with Form PCT/IB/301).

Where a translation of the international application must be furnished to an elected Office, that translation must contain a translation of any annexes to the international preliminary examination report. It is the applicant's responsibility to prepare and furnish such translation directly to each elected Office concerned.

For further details on the applicable time limits and requirements of the elected Offices, see Volume II of the PCT Applicant's Guide.

Name and mailing address of the IPEA/



European Patent Office  
D-80298 Munich  
Tel. +49 89 2399 - 0 Tx: 523656 epmu d  
Fax: +49 89 2399 - 4465

Authorized officer

Neumann, M

Tel. +49 89 2399-7351







## PATENT COOPERATION TREATY

## PCT

## INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

Applicant's or agent's file reference 9579-24		<b>FOR FURTHER ACTION</b> See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)	
International application No. PCT/CA00/00191	International filing date (day/month/year) 25/02/2000	Priority date (day/month/year) 26/02/1999	
International Patent Classification (IPC) or national classification and IPC A61K39/395			
Applicant LEVY, Gary			
<p>1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.</p> <p>2. This REPORT consists of a total of 8 sheets, including this cover sheet.</p> <p><input type="checkbox"/> This report is also accompanied by ANNEXES, i.e. sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).</p> <p>These annexes consist of a total of sheets.</p>			
<p>3. This report contains indications relating to the following items:</p> <ul style="list-style-type: none"> <li>I <input checked="" type="checkbox"/> Basis of the report</li> <li>II <input type="checkbox"/> Priority</li> <li>III <input checked="" type="checkbox"/> Non-establishment of opinion with regard to novelty, inventive step and industrial applicability</li> <li>IV <input type="checkbox"/> Lack of unity of invention</li> <li>V <input checked="" type="checkbox"/> Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement</li> <li>VI <input type="checkbox"/> Certain documents cited</li> <li>VII <input checked="" type="checkbox"/> Certain defects in the international application</li> <li>VIII <input checked="" type="checkbox"/> Certain observations on the international application</li> </ul>			
Date of submission of the demand 18/09/2000		Date of completion of this report 09.05.2001	
Name and mailing address of the international preliminary examining authority:  European Patent Office D-80298 Munich Tel. +49 89 2399 - 0 Tx: 523656 epmu d Fax: +49 89 2399 - 4465		Authorized officer BROCHADO GARGANTA, M Telephone No. +49 89 2399 8935 	



**INTERNATIONAL PRELIMINARY  
EXAMINATION REPORT**

International application No. PCT/CA00/00191

**I. Basis of the report**

1. With regard to the **elements** of the international application (*Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to this report since they do not contain amendments (Rules 70.16 and 70.17)*):

**Description, pages:**

1-45 as originally filed

**Claims, No.:**

1-18 as originally filed

**Drawings, sheets:**

1/13-13/13 as originally filed

**Sequence listing part of the description, pages:**

1-4, filed with the letter of 13.07.2001

2. With regard to the **language**, all the elements marked above were available or furnished to this Authority in the language in which the international application was filed, unless otherwise indicated under this item.

These elements were available or furnished to this Authority in the following language: , which is:

- ☐ the language of a translation furnished for the purposes of the international search (under Rule 23.1(b)).
- ☐ the language of publication of the international application (under Rule 48.3(b)).
- ☐ the language of a translation furnished for the purposes of international preliminary examination (under Rule 55.2 and/or 55.3).

3. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international preliminary examination was carried out on the basis of the sequence listing:

- ☐ contained in the international application in written form.
- ☐ filed together with the international application in computer readable form.
- ☒ furnished subsequently to this Authority in written form.
- ☒ furnished subsequently to this Authority in computer readable form.
- ☒ The statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.
- ☒ The statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished.

4. The amendments have resulted in the cancellation of:





**INTERNATIONAL PRELIMINARY  
EXAMINATION REPORT**

International application No. PCT/CA00/00191

- ☐ the description, pages:
- ☐ the claims, Nos.:
- ☐ the drawings, sheets:

5. ☐ This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)):

*(Any replacement sheet containing such amendments must be referred to under item 1 and annexed to this report.)*

6. Additional observations, if necessary:

**III. Non-establishment of opinion with regard to novelty, inventive step and industrial applicability**

1. The questions whether the claimed invention appears to be novel, to involve an inventive step (to be non-obvious), or to be industrially applicable have not been examined in respect of:

- ☐ the entire international application.
- ☒ claims Nos. 1-6, 11-16.

because:

- ☒ the said international application, or the said claims Nos. 1-6, 11-16, with respect to industrial applicability relate to the following subject matter which does not require an international preliminary examination (*specify*):  
**see separate sheet**

- ☐ the description, claims or drawings (*indicate particular elements below*) or said claims Nos. are so unclear that no meaningful opinion could be formed (*specify*):

- ☐ the claims, or said claims Nos. are so inadequately supported by the description that no meaningful opinion could be formed.

- ☐ no international search report has been established for the said claims Nos. .

2. A meaningful international preliminary examination cannot be carried out due to the failure of the nucleotide and/or amino acid sequence listing to comply with the standard provided for in Annex C of the Administrative Instructions:

- ☐ the written form has not been furnished or does not comply with the standard.
- ☐ the computer readable form has not been furnished or does not comply with the standard.

**V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement**



# **I INTERNATIONAL PRELIMINARY EXAMINATION REPORT**

International application No. PCT/CA00/00191

## **1. Statement**

Novelty (N)	Yes:	Claims	3, 4, 9-18
	No:	Claims	1, 2, 5-8
Inventive step (IS)	Yes:	Claims	
	No:	Claims	1-18
Industrial applicability (IA)	Yes:	Claims	7-10, 17-18
	No:	Claims	

## **2. Citations and explanations see separate sheet**

## **VII. Certain defects in the international application**

The following defects in the form or contents of the international application have been noted:  
**see separate sheet**

## **VIII. Certain observations on the international application**

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:  
**see separate sheet**



**Re Item I**

**Basis of the report**

The Sequence listing pages 1-4 are not considered to be part of the application as they were not filed with the application (Rule 13ter.1-(f) PCT).

**Re Item III**

**Non-establishment of opinion with regard to novelty, inventive step and industrial applicability**

Claims 1-6 and 11-16 are directed to a method of treatment of the human/animal body. They relate therefore to subject-matter considered by this Authority to be covered by the provisions of Rule 67.1(iv) PCT. Consequently, no opinion will be formulated with respect to the industrial applicability of the subject-matter of these claims (Article 34(4)(a)(i) PCT).

**Re Item V**

**Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement**

1. Reference is made to the following documents:

- (B) Ning Q et al.: Hepatology, vol. 28, no. 4 Part 2, October 1998 (1998-10), page 405A
- (C) Stohlman S. et al.: Virology 202, 146-153 (1994)
- (D) Ning Q et al.: Journal of Biological Chemistry, vol. 274, no. 15, 9 April 1999 (1999-04-09), pages 9930-9936
- (E) Levy Gary et al.: Hepatology, vol. 30, no. 4 Part 2, October 1999 (1999-10), page 502A



2. Documents D and E cited in the Search Report as intermediate documents are to be considered as state of the art for claims 11-18, as the date of priority is not validly claimed for these claims.

The administration of an inhibitor of LF-A1 gene or protein or a composition containing it, are not disclosed in the priority document.

3. Novelty

- 3.1 The subject-matter of claims 1, referring to a method for preventing or reducing immune coagulation caused by a virus, is not new in the sense of Article 33(2) PCT, because the method with these features is already disclosed in document A.

Document A discloses a method for inhibiting viral replication, wherein antisense nucleic acid molecules against the nucleocapsid protein gene of mouse hepatitis virus are administered (see abstract).

- 3.2 The same applies to dependent claims 2 and 5. Their features are already known from document A (see abstract).

- 3.3 The subject-matter of claims 6, referring to a method for inducing immune coagulation, is not new in the sense of Article 33(2) PCT, because the method with these features is already known from document B.

Document B discloses a method for inducing immune coagulation, wherein the nucleocapsid (N) protein of murine hepatitis virus type 3 induces transcription of the fgl-2 prothrombinase gene, resulting in an increased functional procoagulant activity (see abstract).

- 3.4 Claims 7 and 8, relate respectively to a composition for use in inhibiting and inducing procoagulant activity. Such compositions are already known from documents A and B (see abstract).

4. Inventive step





- 4.1 Claims 3 and 4 do not contain any additional features which in combination with the features of claim 1, meet the requirements of the PCT in respect of inventive step (Article 33(3) PCT).

The additional features of these claims, i.e. the induction of fgl-2 by the hepatitis virus and having an antibody that binds to the N-protein or to domain I of the N-protein, are well known in the art. The induction of fgl-2 is a functional characteristic inherent of the nucleocapsid protein of hepatitis virus (see for example document B, title). The use of antibodies as inhibitor of protein N will be obvious for the skilled person, as they would have a high affinity for this protein. They are also normally used in the art (see for example document C, abstract).

- 4.2 Claims 9 and 10 relate to a vaccine comprising an effective amount of an N-protein. It is known in the art that hepatitis virus N protein, which forms the viral helical nucleocapsids, performs a number of regulatory functions during virus replication (see for example the abstracts of documents A, B and C). Thus, it would be obvious for the skilled person to use this protein for a vaccine against hepatitis. Thus, claims 9 and 10 are not based on an inventive concept as required by Article 33(3) PCT.
- 4.3 Claim 11 refers to a method for preventing or reducing immune coagulation, wherein an effective amount of an inhibitor of LF-A1 gene or protein is administered.

It is known from document D that there are three putative cis-elements in the fgl2 promoter, that are necessary for transcription. One of these elements is the LF-A1, which is known to regulate gene expression of coagulation factors (see page 9934, column 2). Thus, the skilled person would consider the use of an inhibitor of this element for the method of claim 1, without for that the need of an inventive concept. Therefore, claim 11 is not inventive (Article 33(3) PCT).

- 4.4 Considering the reasoning given in 4.3 and with regard to the reasoning given in this communication for claims 1-10, it is considered that claims 12-18 are also not based on an inventive step as required by Article 33(3) PCT.
5. For the assessment of the present claims 1-6 and 11-16 on the question whether they are industrially applicable, no unified criteria exist in the PCT Contracting States.



The patentability can also be dependent upon the formulation of the claims. The EPO, for example, does not recognize as industrially applicable the subject-matter of claims to the use of a compound in medical treatment, but may allow, however, claims to a known compound for first use in medical treatment and the use of such a compound for the manufacture of a medicament for a new medical treatment.

**Re Item VII**

**Certain defects in the international application**

1. Dependent claim 14 has a back reference to claim 1, but it is more probably depending on claim 11.

**Re Item VIII**

**Certain observations on the international application**

1. The use of general statements in the description which imply that the extent of protection may be expanded in some vague and not precisely defined way ("spirit of the invention" on pages 4, line 20, and 34, line 24), is not allowable (Article 6 PCT, see Guidelines III-4.3.a).



## TENT COOPERATION TREAT

## PCT

## INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference <b>9579-24</b>	<b>FOR FURTHER ACTION</b> see Notification of Transmittal of International Search Report (Form PCT/ISA/220) as well as, where applicable, item 5 below.	
International application No. <b>PCT/CA 00/ 00191</b>	International filing date (day/month/year) <b>25/02/2000</b>	(Earliest) Priority Date (day/month/year) <b>26/02/1999</b>
Applicant  <b>LEVY, Gary</b>		

This International Search Report has been prepared by this International Searching Authority and is transmitted to the applicant according to Article 18. A copy is being transmitted to the International Bureau.

This International Search Report consists of a total of 5 sheets.

☒ It is also accompanied by a copy of each prior art document cited in this report.

**1. Basis of the report**

- a. With regard to the **language**, the international search was carried out on the basis of the international application in the language in which it was filed, unless otherwise indicated under this item.

☐ the international search was carried out on the basis of a translation of the international application furnished to this Authority (Rule 23.1(b)).

- b. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international search was carried out on the basis of the sequence listing :

☐ contained in the international application in written form.

☐ filed together with the international application in computer readable form.

☒ furnished subsequently to this Authority in written form.

☒ furnished subsequently to this Authority in computer readable form.

☒ the statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.

☒ the statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished

2. ☒ **Certain claims were found unsearchable** (See Box I).

3. ☐ **Unity of Invention is lacking** (see Box II).

4. With regard to the **title**,

☐ the text is approved as submitted by the applicant.

☒ the text has been established by this Authority to read as follows:

**MODULATORS OF FGL2 PROTHROMBINASE**

5. With regard to the **abstract**,

☒ the text is approved as submitted by the applicant.

☐ the text has been established, according to Rule 38.2(b), by this Authority as it appears in Box III. The applicant may, within one month from the date of mailing of this international search report, submit comments to this Authority.

6. The figure of the **drawings** to be published with the abstract is Figure No.

☐ as suggested by the applicant.

☐ because the applicant failed to suggest a figure.

☐ because this figure better characterizes the invention.

☒ None of the figures.



## INTERNATIONAL SEARCH REPORT

National Application No.

PCT/CA 00/00191

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 A61K39/395 A61K38/16 C12N15/11 A61K48/00 A61K39/12  
 A61K39/29 A61P7/00 A61P7/02 //C07K16/28

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

MEDLINE, BIOSIS, WPI Data, EPO-Internal, PAJ

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>NING Q ET AL: "The nucleocapsid (N) protein of murine hepatitis virus type 3 (MHV-3) induces transcription of the novel fgl-2 prothrombinase gene."</p> <p>HEPATOLOGY, vol. 28, no. 4 PART 2, October 1998 (1998-10), page 405A XP000929770</p> <p>Biennial Scientific Meeting of the International Association for the Study of the Liver and the 49th Annual Meeting and Postgraduate Courses of the American Association for the Study of Liver Diseases; Chicago, Illinois, USA; November 4-10, 1998 ISSN: 0270-9139 the whole document</p> <p style="text-align: center;">---</p> <p style="text-align: center;">-/--</p>	1-10



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

## \* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

7 August 2000

Date of mailing of the international search report

23/08/2000

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2  
 NL - 2280 HV Rijswijk  
 Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,  
 Fax: (+31-70) 340-3016

Authorized officer

Covone, M





## INTERNATIONAL SEARCH REPORT

International Application No

PCT/CA 00/00191

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>STOHLMAN S A ET AL: "Location of antibody epitopes within the mouse hepatitis virus nucleocapsid protein."            VIROLOGY, (1994 JUL) 202 (1) 146-53. ,            XP002144399            abstract            page 151, right-hand column, line 15 -page 152, left-hand column, line 18            ---</p>	7-10
X	<p>DATABASE MEDLINE 'Online!            US NATIONAL LIBRARY OF MEDICINE (NLM),            BETHESDA, MD, US;            MIZUTANI T ET AL: "Both antisense and sense RNAs against the nucleocapsid protein gene inhibit the multiplication of mouse hepatitis virus."            retrieved from STN            Database accession no. 94355440            XP002144401            abstract            &amp; JOURNAL OF VETERINARY MEDICAL SCIENCE,            (1994 APR) 56 (2) 211-5. ,            ---</p>	7,8
X	<p>US 5 849 485 A (ZHONG WEIMIN ET AL)            15 December 1998 (1998-12-15)            column 1, line 25-39            column 30, line 56 -column 31, line 4            ---</p>	17,18
A	<p>DING J W ET AL: "Cloning and characterization of the human prothrombinase gene (H-fgl2) and its role in human hepatitis."            HEPATOLOGY,            vol. 28, no. 4 PART 2,            October 1998 (1998-10), page 254A            XP000929678            Biennial Scientific Meeting of the International Association for the Study of the Liver and the 49th Annual Meeting and Postgraduate Courses of the American Association for the Study of Liver Diseases; Chicago, Illinois, USA; November 4-10, 1998            ISSN: 0270-9139            the whole document            ---            -/--</p>	1-18



## INTERNATIONAL SEARCH REPORT

International Application No.

PCT/CA 00/00191

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P, X	NING QIN ET AL: "The nucleocapsid protein of murine hepatitis virus type 3 induces transcription of the novel fgl2 prothrombinase gene." JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 274, no. 15, 9 April 1999 (1999-04-09), pages 9930-9936, XP002144400 ISSN: 0021-9258 the whole document -----	1-18
P, X	LEVY GARY A ET AL: "Study of putative CIS-elements in MFGL2 promoter in response to nucleocapsid protein of murine hepatitis virus type 3." HEPATOLOGY, vol. 30, no. 4 PART 2, October 1999 (1999-10), page 502A XP000929772 50th Annual Meeting and Postgraduate Courses of the American Association for the Study of Liver Diseases; Dallas, Texas, USA; November 5-9, 1999 ISSN: 0270-9139 the whole document -----	1-18



# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/CA 00/00191

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US 5849485 A	15-12-1998	US 5604115 A	18-02-1997
		US 6025196 A	15-02-2000
		AT 185598 T	15-10-1999
		AU 665939 B	25-01-1996
		AU 9174291 A	22-07-1992
		CA 2098838 A	22-06-1992
		DE 69131718 D	18-11-1999
		DE 69131718 T	02-03-2000
		EP 0564592 A	13-10-1993
		JP 6505152 T	16-06-1994
		WO 9211365 A	09-07-1992
<hr/>			



# INTERNATIONAL ARCH REPORT

International Application No

PCT/CA 00/00191

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 A61K39/395 A61K38/16 C12N15/11 A61K48/00 A61K39/12  
A61K39/29 A61P7/00 A61P7/02 //C07K16/28

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

MEDLINE, BIOSIS, WPI Data, EPO-Internal, PAJ

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>NING Q ET AL: "The nucleocapsid (N) protein of murine hepatitis virus type 3 (MHV-3) induces transcription of the novel fgl-2 prothrombinase gene."</p> <p>HEPATOLOGY, vol. 28, no. 4 PART 2, October 1998 (1998-10), page 405A</p> <p>XP000929770</p> <p>Biennial Scientific Meeting of the International Association for the Study of the Liver and the 49th Annual Meeting and Postgraduate Courses of the American Association for the Study of Liver Diseases; Chicago, Illinois, USA; November 4-10, 1998</p> <p>ISSN: 0270-9139</p> <p>the whole document</p> <p style="text-align: center;">---</p> <p style="text-align: center;">-/--</p>	1-10

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

### \* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

7 August 2000

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